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(54) BMP-9 COMPOSITIONS

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COMPOSITIONS BMP-9

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CELESTE, A.J. ET AL. 'Identification of transforming growth factor beta family members present in bone-inductive protein purified from bovine bone'

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Description

The present invention relates to a novel family of purified proteins designated BMP-9 proteins and processes for obtaining them. These proteins may be used to induce bone and/or cartilage formation and in wound healing and tissue repair.

The murine BMP-9 DNA sequence (SEQ ID NO: 1) and amino acid sequence (SEQ ID NO: 2) are set forth in Figure 1. Human BMP-9 sequence is set forth in Figure 3 (SEQ ID NO: 8 and SEQ ID NO: 9). It is contemplated that BMP-9 proteins are capable of inducing the formation of cartilage and/or bone. BMP-9 proteins may be further characterized by the ability to demonstrate cartilage and/or bone formation activity in the rat bone formation assay described below.

Murine BMP-9 is characterized by comprising amino acid #319 to #428 of Figure 1 (SEQ ID NO: 2 amino acids #1-110). Murine BMP-9 may be produced by culturing a cell transformed with a DNA sequence comprising nucleotide #610 to nucleotide #1893 as shown in Figure 1 (SEQ ID NO: 1) and recovering and purifying from the culture medium a protein characterized by the amino acid sequence comprising amino acid #319 to #428 as shown in Figure 1 (SEQ ID NO: 2) substantially free from other proteinaceous materials with which it is co-produced.

Human BMP-9 is expected to be homologous to murine BMP-9 and is characterized by comprising amino acid #1 (Ser, Ala, Gly) to #110 of Figure 3 (SEQ ID NO: 9) (Arg). The invention includes methods for obtaining the DNA sequences encoding human BMP-9. This method entails utilizing the murine BMP-9 nucleotide sequence or portions thereof to design probes to screen libraries for the human gene or fragments thereof using standard techniques. Human BMP-9 may be produced by culturing a cell transformed with the BMP-9 DNA sequence and recovering and purifying BMP-9 from the culture medium. The expressed protein is isolated, recovered, and purified from the culture medium. The purified expressed protein is substantially free from other proteinaceous materials with which it is co-produced, as well as from other contaminants. The recovered purified protein is contemplated to exhibit cartilage and/or bone formation activity. The proteins of the invention may be further characterized by the ability to demonstrate cartilage and/or bone formation activity in the rat bone formation assay described below.

Human BMP-9 may be produced by culturing a cell transformed with a DNA sequence comprising nucleotide #124 to #453 as shown in SEQ ID NO: 8 and recovering and purifying from the culture medium a protein characterized by the amino acid sequence of SEQ ID NO: 9 from amino acid #1 to amino acid #110 substantially free from other proteinaceous materials with which it is co-produced.

Another aspect of the invention provides pharmaceutical compositions containing a therapeutically effective amount of a BMP-9 protein in a pharmaceutically acceptable vehicle or carrier. BMP-9 compositions of the invention may be used in the formation of cartilage. These compositions may further be utilized for the formation of bone. BMP-9 compositions may also be used for wound healing and tissue repair. Compositions of the invention may further include at least one other therapeutically useful agent such as the BMP proteins BMP-1, BMP-2, BMP-3, BMP-4, BMP-5, BMP-6, and BMP-7 disclosed for instance in PCT publications W088/00205, W089/10409, and W090/11366, and BMP-8, disclosed in U.S. application Ser. No. 07/641,204 filed January 15, 1991, Ser. No. 07/525,357 filed May 16, 1990, and Ser. No. 07/800,364 filed November 20, 1991.

The compositions of the invention may comprise, in addition to a BMP-9 protein, other therapeutically useful agents including growth factors such as epidermal growth factor (EGF), fibroblast growth factor (FGF), transforming growth factor (TGF- α and TGF- β), and insulin-like growth factor (IGF). The compositions may also include an appropriate matrix for instance, for supporting the composition and providing a surface for bone and/or cartilage growth. The matrix may provide slow release of the osteoinductive protein and/or the appropriate environment for presentation thereof.

The BMP-9 compositions may be employed in methods for treating a number of bone and/or cartilage defects, periodontal disease and various types of wounds. These methods, according to the invention, entail administering to a patient needing such bone and/or cartilage formation wound healing or tissue repair, an effective amount of a BMP-9 protein. These methods may also entail the administration of a protein of the invention in conjunction with at least one of the novel BMP proteins disclosed in the co-owned applications described above. In addition, these methods may also include the administration of a BMP-9 protein with other growth factors including EGF, FGF, TGF- α , TGF- β , and IGF.

Still a further aspect of the invention are DNA sequences coding for expression of a BMP-9 protein. Such sequences include the sequence of nucleotides in a 5' to 3' direction illustrated in Figure 1 (SEQ ID NO: 1) and Figure 3 (SEQ ID NO: 8) or DNA sequences which hybridize under stringent conditions with the DNA sequences of Figure 1 or 3 and encode a protein having the ability to induce the formation of cartilage and/or bone. Finally, allelic or other variations of the sequences of Figure 1 or 3, whether such nucleotide changes result in changes in the peptide sequence or not, are also included in the present invention.

A further aspect of the invention includes vectors comprising a DNA sequence as described above in operative association with an expression control sequence therefor. These vectors may be employed in a novel process for producing a BMP-9 protein of the invention in which a cell line transformed with a DNA sequence encoding a BMP-9

protein in operative association with an expression control sequence therefor, is cultured in a suitable culture medium and a BMP-9 protein is recovered and purified therefrom. This process may employ a number of known cells both prokaryotic and eukaryotic as host cells for expression of the polypeptide.

Other aspects and advantages of the present invention will be apparent upon consideration of the following detailed description and preferred embodiments thereof.

Brief Description of the Drawings

FIG. 1 comprises DNA sequence and derived amino acid sequence of murine BMP-9 from clone ML14a further described below.

FIG. 2 comprises DNA sequence and derived amino acid sequence of human BMP-4 from lambda U20S-3 ATCC #40342.

FIG. 3 comprises DNA sequence and derived amino acid sequence of human BMP-9 from λ FIX/H6III ATCC #75252.

Detailed Description of the Invention

The murine BMP-9 nucleotide sequence (SEQ ID NO: 1) and encoded amino acid sequence (SEQ ID NO: 2) are depicted in Figure 1. Purified murine BMP-9 proteins of the present invention are produced by culturing a host cell transformed with a DNA sequence comprising the DNA coding sequence of Figure 1 (SEQ ID NO: 1) from nucleotide #610 to nucleotide #1893 and recovering and purifying from the culture medium a protein which contains the amino acid sequence or a substantially homologous sequence as represented by amino acid #319 to #428 of Figure 1 (SEQ ID NO: 2). The BMP-9 proteins recovered from the culture medium are purified by isolating them from other proteinaceous materials from which they are co-produced and from other contaminants present.

Human BMP-9 nucleotide and amino acid sequence is depicted in SEQ ID No: 8 and 9. Mature human BMP-9 is expected to comprise amino acid #1 (Ser, Ala, Gly) to #110 (Arg).

Human BMP-9 may be produced by culturing a cell transformed with a DNA sequence comprising nucleotide #124 to #453 as shown in SEQ ID NO: 8 and recovering and purifying from the culture medium a protein characterized by the amino acid sequence of SEQ ID NO: 9 from amino acid #1 to amino acid #110 substantially free from other proteinaceous materials with which it is co-produced.

BMP-9 proteins may be characterized by the ability to induce the formation of cartilage. BMP-9 proteins may be further characterized by the ability to induce the formation of bone. BMP-9 proteins may be further characterized by the ability to demonstrate cartilage and/or bone formation activity in the rat bone formation assay described below.

The BMP-9 proteins provided herein also include factors encoded by sequences similar to those of Figure 1 and 3 (SEQ ID NO's: 1 and 8), but into which modifications are naturally provided (e.g. allelic variations in the nucleotide sequence which may result in amino acid changes in the polypeptide) or deliberately engineered. For example, synthetic polypeptides may wholly or partially duplicate continuous sequences of the amino acid residues of Figure 1 of Figure 3 (SEQ ID NO's: 2 and 9). These sequences, by virtue of sharing primary, secondary, or tertiary structural and conformational characteristics with bone growth factor polypeptides of Figure 1 and Figure 3 may possess bone growth factor biological properties in common therewith. Thus, they may be employed as biologically active substitutes for naturally-occurring BMP-9 and other BMP-9 polypeptides in therapeutic processes.

Other specific mutations of the sequences of BMP-9 proteins described herein involve modifications of glycosylation sites. These modifications may involve O-linked or N-linked glycosylation sites. For instance, the absence of glycosylation or only partial glycosylation results from amino acid substitution or deletion at asparagine-linked glycosylation recognition sites. The asparagine-linked glycosylation recognition sites comprise tripeptide sequences which are specifically recognized by appropriate cellular glycosylation enzymes. These tripeptide sequences are either asparagine-X-threonine or asparagine-X-serine, where X is usually any amino acid. A variety of amino acid substitutions or deletions at one or both of the first or third amino acid positions of a glycosylation recognition site (and/or amino acid deletion at the second position) results in non-glycosylation at the modified tripeptide sequence.

The present invention also encompasses the novel DNA sequences, free of association with DNA sequences encoding other proteinaceous materials, and coding on expression for BMP-9 proteins. These DNA sequences include those depicted in Figure 1 or Figure 3 (SEQ ID NO's: 1 and 8) in a 5' to 3' direction and those sequences which hybridize thereto under stringent hybridization conditions [see, T. Maniatis et al, Molecular Cloning (A Laboratory Manual), Cold Spring Harbor Laboratory (1982), pages 387 to 389] and encode a protein having cartilage and/or bone inducing activity.

Similarly, DNA sequences which code for BMP-9 proteins coded for by the sequences of Figure 1 or Figure 3, but which differ in codon sequence due to the degeneracies of the genetic code or allelic variations (naturally-occurring base changes in the species population which may or may not result in an amino acid change) also encode the novel factors described herein. Variations in the DNA sequences of Figure 1 or Figure 3 (SEQ ID NO: 1 and 8) which are

caused by point mutations or by induced modifications (including insertion, deletion, and substitution) to enhance the activity, half-life or production of the polypeptides encoded are also encompassed in the invention.

Another aspect of the present invention provides a novel method for producing BMP-9 proteins. The method of the present invention involves culturing a suitable cell line, which has been transformed with a DNA sequence encoding a BMP-9 protein of the invention, under the control of known regulatory sequences. The transformed host cells are cultured and the BMP-9 proteins recovered and purified from the culture medium. The purified proteins are substantially free from other proteins with which they are co-produced as well as from other contaminants.

Suitable cells or cell lines may be mammalian cells, such as Chinese hamster ovary cells (CHO). The selection of suitable mammalian host cells and methods for transformation, culture, amplification, screening, product production and purification are known in the art. See, e.g., Gething and Sambrook, *Nature*, 293:620-625 (1981), or alternatively, Kaufman et al, *Mol. Cell. Biol.*, 5(7):1750-1759 (1985) or Howley et al, U.S. Patent 4,419,446. Another suitable mammalian cell line, which is described in the accompanying examples, is the monkey COS-1 cell line. The mammalian cell CV-1 may also be suitable.

Bacterial cells may also be suitable hosts. For example, the various strains of *E. coli* (e.g., HB101, MCI061) are well-known as host cells in the field of biotechnology. Various strains of *B. subtilis*, *Pseudomonas*, other bacilli and the like may also be employed in this method.

Many strains of yeast cells known to those skilled in the art may also be available as host cells for expression of the polypeptides of the present invention. Additionally, where desired, insect cells may be utilized as host cells in the method of the present invention. See, e.g. Miller et al, *Genetic Engineering*, 8:277-298 (Plenum Press 1986) and references cited therein.

Another aspect of the present invention provides vectors for use in the method of expression of these novel BMP-9 polypeptides. Preferably the vectors contain the full novel DNA sequences described above which encode the novel factors of the invention. Additionally the vectors also contain appropriate expression control sequences permitting expression of the BMP-9 protein sequences. Alternatively, vectors incorporating modified sequences as described above are also embodiments of the present invention. The vectors may be employed in the method of transforming cell lines and contain selected regulatory sequences in operative association with the DNA coding sequences of the invention which are capable of directing the replication and expression thereof in selected host cells. Regulatory sequences for such vectors are known to those skilled in the art and may be selected depending upon the host cells. Such selection is routine and does not form part of the present invention.

A protein of the present invention, which induces cartilage and/or bone formation in circumstances where bone is not normally formed, has application in the healing of bone fractures and cartilage defects in humans and other animals. Such a preparation employing a BMP-9 protein may have prophylactic use in closed as well as open fracture reduction and also in the improved fixation of artificial joints. De novo bone formation induced by an osteogenic agent contributes to the repair of congenital, trauma induced, or oncologic resection induced craniofacial defects, and also is useful in cosmetic plastic surgery. A BMP-9 protein may be used in the treatment of periodontal disease, and in other tooth repair processes. Such agents may provide an environment to attract bone-forming cells, stimulate growth of bone-forming cells or induce differentiation of progenitors of bone-forming cells. BMP-9 polypeptides of the invention may also be useful in the treatment of osteoporosis. A variety of osteogenic, cartilage-inducing and bone inducing factors have been described. See, e.g. European patent applications 148,155 and 169,016 for discussions thereof.

The proteins of the invention may also be used in wound healing and related tissue repair. The types of wounds include, but are not limited to burns, incisions and ulcers. (See, e.g. PCT Publication W084/01106 for discussion of wound healing and related tissue repair).

It is further contemplated that proteins of the invention may increase neuronal survival and therefore be useful in transplantation and treatment of conditions exhibiting a decrease in neuronal survival.

A further aspect of the invention is a therapeutic method and composition for repairing fractures and other conditions related to cartilage and/or bone defects or periodontal diseases. The invention further comprises therapeutic methods and compositions for wound healing and tissue repair. Such compositions comprise a therapeutically effective amount of at least one of the BMP-9 proteins of the invention in admixture with a pharmaceutically acceptable vehicle, carrier or matrix.

It is expected that the proteins of the invention may act in concert with or perhaps synergistically with other related proteins and growth factors. Further therapeutic methods and compositions of the invention therefore comprise a therapeutic amount of at least one BMP-9 protein of the invention with a therapeutic amount of at least one of the other BMP proteins disclosed in co-owned applications described above. Such combinations may comprise separate molecules of the BMP proteins or heteromolecules comprised of different BMP moieties. For example, a method and composition of the invention may comprise a disulfide linked dimer comprising a BMP-9 protein subunit and a subunit from one of the "BMP" proteins described above. A further embodiment may comprise a heterodimer of BMP-9 moieties. Further, BMP-9 proteins may be combined with other agents beneficial to the treatment of the bone and/or cartilage defect, wound, or tissue in question. These agents include various growth factors such as epidermal growth factor

(EGF), platelet derived growth factor (PDGF), transforming growth factors (TGF- α and TGF- β), and insulin-like growth factor (IGF).

The preparation and formulation of such physiologically acceptable protein compositions, having due regard to pH, isotonicity, stability and the like, is within the skill of the art. The therapeutic compositions are also presently valuable for veterinary applications due to the lack of species specificity in BMP proteins. Particularly domestic animals and thoroughbred horses in addition to humans are desired patients for such treatment with BMP-9 of the present invention.

The therapeutic method includes administering the composition topically, systemically, or locally as an implant or device. When administered, the therapeutic composition for use in this invention is, of course, in a pyrogen-free, physiologically acceptable form. Further, the composition may desirably be encapsulated or injected in a viscous form for delivery to the site of bone, cartilage or tissue damage. Topical administration may be suitable for wound healing and tissue repair. Therapeutically useful agents other than the BMP-9 proteins which may also optionally be included in the composition as described above, may alternatively or additionally, be administered simultaneously or sequentially with the BMP composition in the methods of the invention.

Preferably for bone and/or cartilage formation, the composition would include a matrix capable of delivering BMP-9 or other BMP proteins to the site of bone and/or cartilage damage, providing a structure for the developing bone and cartilage and optimally capable of being resorbed into the body. The matrix may provide slow release of BMP-9 and/or the appropriate environment for presentation thereof. Such matrices may be formed of materials presently in use for other implanted medical applications.

The choice of matrix material is based on biocompatibility, biodegradability, mechanical properties, cosmetic appearance and interface properties. The particular application of the BMP-9 compositions will define the appropriate formulation. Potential matrices for the compositions may be biodegradable and chemically defined calcium sulfate, tricalciumphosphate, hydroxyapatite, polylactic acid and polyanhydrides. Other potential materials are biodegradable and biologically well defined, such as bone or dermal collagen. Further matrices are comprised of pure proteins or extracellular matrix components. Other potential matrices are nonbiodegradable and chemically defined, such as sintered hydroxyapatite, bioglass, aluminates, or other ceramics. Matrices may be comprised of combinations of any of the above mentioned types of material, such as polylactic acid and hydroxyapatite or collagen and tricalciumphosphate. The bioceramics may be altered in composition, such as in calcium-aluminate-phosphate and processing to alter pore size, particle size, particle shape, and biodegradability.

The dosage regimen will be determined by the attending physician considering various factors which modify the action of the BMP-9 protein, e.g. amount of bone weight desired to be formed, the site of bone damage, the condition of the damaged bone, the size of a wound, type of damaged tissue, the patient's age, sex, and diet, the severity of any infection, time of administration and other clinical factors. The dosage may vary with the type of matrix used in the reconstitution and the types of BMP proteins in the composition. The addition of other known growth factors, such as IGF I (insulin like growth factor I), to the final composition, may also effect the dosage. Progress can be monitored by periodic assessment of bone growth and/or repair, for example, x-rays, histomorphometric determinations and tetracycline labeling.

The following examples illustrate practice of the present invention in recovering and characterizing murine BMP-9 protein and employing it to recover the human and other BMP-9 proteins, obtaining the human proteins and expressing the proteins via recombinant techniques.

EXAMPLE I

Murine BMP-9

750,000 recombinants of a mouse liver cDNA library made in the vector lambdaZAP (Stratagene/Catalog #935302) are plated and duplicate nitrocellulose replicas made. A fragment of human BMP-4 DNA corresponding to nucleotides 1330-1627 of Figure 2 (SEQ ID NO: 3) (the human BMP-4 sequence) is ^{32}P -labeled by the random priming procedure of Feinberg et al. [Anal. Biochem. 132: 6-13 (1983)] and hybridized to both sets of filters in SHB at 60°C for 2 to 3 days. Both sets of filters are washed under reduced stringency conditions (4X SSC, 0.1% SDS at 60°C). Many duplicate hybridizing recombinants of various intensities (approximately 92) are noted. 50 of the strongest hybridizing recombinant bacteriophage are plaque purified and their inserts are transferred to the plasmid Bluescript SK (+/-) according to the *in vivo* excision protocol described by the manufacturer (Stratagene). DNA sequence analysis of several recombinants indicate that they encode a protein homologous to other BMP proteins and other proteins in the TGF- β family. The DNA sequence and derived amino acid sequence of one recombinant, designated ML14a, is set forth in Figure 1. (SEQ ID NO: 1)

The nucleotide sequence of clone ML14a contains an open reading frame of 1284 bp, encoding a BMP-9 protein of 428 amino acids. The encoded 428 amino acid BMP-9 protein is contemplated to be the primary translation product as the coding sequence is preceded by 609 bp of 5' untranslated sequence with stop codons in all three reading frames.

The 428 amino acid sequence predicts a BMP-9 protein with a molecular weight of 48,000 daltons.

Based on knowledge of other BMP proteins and other proteins within the TGF- β family, it is predicted that the precursor polypeptide would be cleaved at the multibasic sequence ARG-ARG-LYS-ARG in agreement with a proposed consensus proteolytic processing sequence of ARG-X-X-ARG. Cleavage of the BMP-9 precursor polypeptide at this location would generate a 110 amino acid mature peptide beginning with the amino acid SER at position #319. The processing of BMP-9 into the mature form is expected to involve dimerization and removal of the N-terminal region in a manner analogous to the processing of the related protein TGF- β [L.E. Gentry, et al., *Molec. & Cell. Biol.* 8:4162 (1988); R. Derynck, et al., *Nature* 316:701 (1985)].

It is contemplated therefore that the mature active species of murine BMP-9 comprises a homodimer of 2 polypeptide subunits, each subunit comprising amino acids #319-#428 with a predicted molecular weight of approximately 12,000 daltons. Further active species are contemplated comprising amino acids #326 - #428 thereby including the first conserved cysteine residue. As with other members of the BMP and TGF- β family of proteins, the carboxy-terminal region of the BMP-9 protein exhibits greater sequence conservation than the more amino-terminal portion. The percent amino acid identity of the murine BMP-9 protein in the cysteine-rich C-terminal domain (amino acids #326 - #428) to the corresponding region of other human BMP proteins and other proteins within the TGF- β family is as follows: BMP-2, 53%; BMP-3, 43%; BMP-4, 53%; BMP-5, 55%; BMP-6, 55%; BMP-7, 53%; Vgl, 50%; GDF-1, 43%; TGF- β 1, 32%; TGF- β 2, 34%; TGF- β 3, 34%; inhibin β (B), 34%; and inhibin β (A), 42%.

EXAMPLE II

Human BMP-9

Murine and human osteoinductive factor genes are presumed to be significantly homologous, therefore the murine coding sequence or a portion thereof is used as a probe to screen a human genomic library or as a probe to identify a human cell line or tissue which synthesizes the analogous human cartilage and/or bone protein. A human genomic library (Toole et al., *supra*) may be screened with such a probe, and presumptive positives isolated and DNA sequence obtained. Evidence that this recombinant encodes a portion of the human BMP-9 relies of the murine/human protein and gene structure homologies.

Once a recombinant bacteriophage containing DNA encoding portion of the human cartilage and/or bone inductive factor molecule is obtained, the human coding sequence can be used as a probe to identify a human cell line or tissue which synthesizes BMP-9. Alternatively, the murine coding sequence can be used as a probe to identify such human cell line or tissue. Briefly described, RNA is extracted from a selected cell or tissue source and either electrophoresed on a formaldehyde agarose gel and transferred to nitrocellulose, or reacted with formaldehyde and spotted on nitrocellulose directly. The nitrocellulose is then hybridized to a probe derived from a coding sequence of the murine or human BMP-9. mRNA is selected by oligo (dT) cellulose chromatography and cDNA is synthesized and cloned in lambda gt10 or lambda ZAP by established techniques (Toole et al., *supra*).

Additional methods known to those skilled in the art may be used to isolate the human and other species' BMP-9 proteins of the invention.

A. Isolation of Human BMP-9 DNA

One million recombinants of a human genomic library constructed in the vector λ FIX (Stratagene catalog # 944201) are plated and duplicate nitrocellulose replicas made. Two oligonucleotides probes designed on the basis of nucleotides #1665-#1704 and #1837-#1876 of the sequence set forth in Figure 1 (SEQ ID NO:1) are synthesized on an automated DNA synthesizer. The sequence of these two oligonucleotides is indicated below:

1: CTATGAGTGTAAGGGGGTTGCTTCTTCCCATTTGGCTGAT

2: GTGCCAACCTCAAGTACCACTATGAGGGGATGAGTGTGG

These two oligonucleotide probes are radioactively labeled with $\gamma^{32}\text{P}$ -ATP and each is hybridized to one set of the duplicate nitrocellulose replicas in SHB at 65°C and washed with 1X SSC, 0.1% SDS at 65°C. Three recombinants which hybridize to both oligonucleotide probes are noted. All three positively hybridizing recombinants are plaque purified, bacteriophage plate stocks are prepared and bacteriophage DNA is isolated from each. The oligonucleotide hybridizing regions of one of these recombinants, designated HGIII, is localized to a 1.2 kb Pst I/Xba I fragment. This fragment is subcloned into a plasmid vector (pGEM-3) and DNA sequence analysis is performed. HGIII was deposited

with the ATCC, 12301 Parklawn Drive, Rockville, Maryland USA on June 16, 1992 under the requirements of the Budapest Treaty and designated as ATCC # 75252. This subclone is designated pGEM-111. A portion of the DNA sequence of clone pGEM-111 is set forth in Figure 3 (SEQ ID NO:8/ HUMAN BMP-9 sequence). This sequence encodes the entire mature region of human BMP-9 and a portion of the propeptide. It should be noted that this sequence consists of preliminary data. Particularly, the propeptide region is subject to further analysis and characterization. For example, nucleotides #1 through #3 (TGA) encode a translational stop which may be incorrect due to the preliminary nature of the sequence. It is predicted that additional sequences present in both pGEM-111 (the 1.2 kb PstI/XbaI fragment of HGIII subcloned into pGEM) and HGIII encode additional amino acids of the human BMP-9 propeptide region. Based on knowledge of other BMPs and other proteins within the TGF- β family, it is predicted that the precursor polypeptide would be cleaved at the multibasic sequence ARG-ARG-LYS-ARG (amino acids # -4 through # -1 of SEQUENCE ID NO:9) in agreement with a proposed consensus proteolytic processing sequence ARG-X-X-ARG. Cleavage of the human BMP-9 precursor polypeptide at this location would generate a 110 amino acid mature peptide beginning with the amino acid SER at position #1 of SEQUENCE ID NO:9 (encoded by nucleotides #124 through #126 of SEQUENCE ID NO:8). The processing of human BMP-9 into the mature form is expected to involve dimerization and removal of the N-terminal region in a manner analogous to the processing of the related protein TGF- β [L.E. Gentry, et al., *Molec. & Cell. Biol.* 8:4162 (1988); R. Derynck, et al., *Nature* 316:701 (1985)].

It is contemplated therefore that the mature active species of human BMP-9 comprises a homodimer of two polypeptide subunits, each subunit comprising amino acids #1 through #110 of SEQUENCE ID NO:9, with a predicted molecular weight of 12,000 daltons. Further active species are contemplated comprising amino acids #8 through #110 thereby including the first conserved cysteine residue. As with other members of the BMP and TGF- β family of proteins, the carboxy-terminal portion of the human BMP-9 sequence exhibits greater sequence conservation than the amino-terminal portion. The percent amino acid identity of the human BMP-9 protein in the cysteine-rich C-terminal domain (amino acids #8 through #110) to the corresponding region of other human BMP proteins and other proteins within the TGF- β family is as follows: BMP-2, 52%; BMP-3, 40%; BMP-4, 52%; BMP-5, 55%; BMP-6, 55%; BMP-7, 53%; murine BMP-9, 97%; Vgl, 50%; GDF-1, 44%; TGF- β 1, 32%; TGF- β 2, 32%; TGF- β 3, 32%; inhibin β (B), 35%; and inhibin β (A), 41%.

EXAMPLE III

Rosen Modified Sampath-Reddi Assay

A modified version of the rat bone formation assay described in Sampath and Reddi, *Proc. Natl. Acad. Sci. U.S.A.*, 80:6591-6595 (1983) is used to evaluate bone and/or cartilage activity of the BMP proteins. This modified assay is herein called the Rosen-modified Sampath-Reddi assay. The ethanol precipitation step of the Sampath-Reddi procedure is replaced by dialyzing (if the composition is a solution) or diafiltering (if the composition is a suspension) the fraction to be assayed against water. The solution or suspension is then redissolved in 0.1 % TFA, and the resulting solution added to 20mg of rat matrix. A mock rat matrix sample not treated with the protein serves as a control. This material is frozen and lyophilized and the resulting powder enclosed in #5 gelatin capsules. The capsules are implanted subcutaneously in the abdominal thoracic area of 21 - 49 day old male Long Evans rats. The implants are removed after 7 - 14 days. Half of each implant is used for alkaline phosphatase analysis [See, A. H. Reddi et al., *Proc. Natl. Acad. Sci.*, 69:1601 (1972)].

The other half of each implant is fixed and processed for histological analysis. 1 μ m glycolmethacrylate sections are stained with Von Kossa and acid fuchsin to score the amount of induced bone and cartilage formation present in each implant. The terms +1 through +5 represent the area of each histological section of an implant occupied by new bone and/or cartilage cells and matrix. A score of +5 indicates that greater than 50% of the implant is new bone and/or cartilage produced as a direct result of protein in the implant. A score of +4, +3, +2 and +1 would indicate that greater than 40%, 30%, 20% and 10% respectively of the implant contains new cartilage and/or bone. In a modified scoring method, three non-adjacent sections are evaluated from each implant and averaged. "+/-" indicates tentative identification of cartilage or bone; "+1" indicates >10% of each section being new cartilage or bone; "+2", >25%; "+3", >50%; "+4", >75%; "+5", >80%. A "-" indicates that the implant is not recovered.

It is contemplated that the dose response nature of the BMP-9 containing samples of the matrix samples will demonstrate that the amount of bone and/or cartilage formed increases with the amount of BMP-9 in the sample. It is contemplated that the control samples will not result in any bone and/or cartilage formation.

As with other cartilage and/or bone inductive proteins such as the above-mentioned "BMP" proteins, the bone and/or cartilage formed is expected to be physically confined to the space occupied by the matrix. Samples are also analyzed by SDS gel electrophoresis and isoelectric focusing followed by autoradiography. The activity is correlated with the protein bands and pl. To estimate the purity of the protein in a particular fraction an extinction coefficient of 1 OD/mg-cm is used as an estimate for protein and the protein is run on SDS PAGE followed by silver staining or radioiodination

5 Expression of BMP-9

One skilled in the art can construct mammalian expression vectors by employing the sequence of Figure 1 (SEQ ID NO: 1) or Figure 3 (SEQ ID NO: 8), or other DNA sequences encoding BMP-9 proteins or other modified sequences and known vectors, such as pCD [Okayama et al., Mol. Cell Biol., 2:161-170 (1982)], pJL3, pJL4 [Gough et al., EMBO J., 4:645-653 (1985)] and pMT2 CXM.

Plasmid pMT2 CXM is obtained by EcoRI digestion of pMT2-VWF, which has been deposited with the American Type Culture Collection (ATCC), Rockville, MD (USA) under accession number ATCC 67122. EcoRI digestion excises the cDNA insert present in pMT2-VWF, yielding pMT2 in linear form which can be ligated and used to transform *E. coli* HB 101 or DH-5 to ampicillin resistance. Plasmid pMT2 DNA can be prepared by conventional methods. pMT2 CXM is then constructed using loopout/in mutagenesis [Morinaga, et al., *Biotechnology* **84**: 636 (1984). This removes bases 1075 to 1145 relative to the Hind III site near the SV40 origin of replication and enhancer sequences of pMT2. In addition it inserts the following sequence:

at nucleotide 1145. This sequence contains the recognition site for the restriction endonuclease Xho I. A derivative of
35 pMT2CXM, termed pMT23, contains recognition sites for the restriction endonucleases PstI, Eco RI, Sall and XhoI.
Plasmid pMT2 CXM and pMT23 DNA may be prepared by conventional methods.

pEMC2bl derived from pMT21 may also be suitable in practice of the invention. pMT21 is derived from pMT2 which is derived from pMT2-VWF. As described above EcoRI digestion excises the cDNA insert present in pMT-VWF, yielding pMT2 in linear form which can be ligated and used to transform E. Coli HB 101 or DH-5 to ampicillin resistance. Plasmid pMT2 DNA can be prepared by conventional methods.

pMT21 is derived from pMT2 through the following two modifications. First, 76 bp of the 5' untranslated region of the DHFR cDNA including a stretch of 19 G residues from G/C tailing for cDNA cloning is deleted. In this process, a XhoI site is inserted to obtain the following sequence immediately

45 upstream from DHFR: 5' -CTGCAGGCGAGCCTGAATTCCTCGAGCCATCATG-3'
PstI Eco RI XhoI
(SEQ ID NO: 6)

50 Second, a unique ClaI site is introduced by digestion with EcoRV and XbaI, treatment with Klenow fragment of DNA polymerase I, and ligation to a ClaI linker (CATCGATG). This deletes a 250 bp segment from the adenovirus associated RNA (VAI) region but does not interfere with VAI RNA gene expression or function. pMT21 is digested with EcoRI and XhoI, and used to derive the vector pEMC2B1.

8

5'-CGAGGTTAAAAACGTCTAGGCCCCCGAACACGGGGACGTGGTTTTTCCTTT
TaqI

5 GAAAAACACGATTGC-3'
XhoI (SEQ ID NO: 7)

10 This sequence matches the EMC virus leader sequence from nucleotide 763 to 827. It also changes the ATG at position 10 within the EMC virus leader to an ATT and is followed by a XhoI site. A three way ligation of the pMT21 Eco RI-XhoI fragment, the EMC virus EcoRI-TaqI fragment, and the 68 bp oligonucleotide adapter TaqI-XhoI adapter resulting in the vector pEMC281.

15 This vector contains the SV40 origin of replication and enhancer, the adenovirus major late promoter, a cDNA copy of the majority of the adenovirus tripartite leader sequence, a small hybrid intervening sequence, an SV40 polyadenylation signal and the adenovirus VA I gene, DHFR and β -lactamase markers and an EMC sequence, in appropriate relationships to direct the high level expression of the desired cDNA in mammalian cells.

20 The construction of vectors may involve modification of the BMP-9 DNA sequences. For instance, BMP-9 cDNA can be modified by removing the non-coding nucleotides on the 5' and 3' ends of the coding region. The deleted non-coding nucleotides may or may not be replaced by other sequences known to be beneficial for expression. These vectors are transformed into appropriate host cells for expression of BMP-9 proteins.

25 One skilled in the art can manipulate the sequences of Figure 1 or Figure 3 (SEQ ID NO: 1 and 8) by eliminating or replacing the mammalian regulatory sequences flanking the coding sequence with bacterial sequences to create bacterial vectors for intracellular or extracellular expression by bacterial cells. For example, the coding sequences could be further manipulated (e.g. ligated to other known linkers or modified by deleting non-coding sequences therefrom or altering nucleotides therein by other known techniques). The modified BMP-9 coding sequence could then be inserted into a known bacterial vector using procedures such as described in T. Taniguchi et al., Proc. Natl Acad. Sci. USA, 77:5230-5233 (1980). This exemplary bacterial vector could then be transformed into bacterial host cells and a BMP-9 protein expressed thereby. For a strategy for producing extracellular expression of BMP-9 proteins in bacterial cells, see, e.g. European patent application EPA 177,343.

30 Similar manipulations can be performed for the construction of an insect vector [See, e.g. procedures described in published European patent application 155,476] for expression in insect cells. A yeast vector could also be constructed employing yeast regulatory sequences for intracellular or extracellular expression of the factors of the present invention by yeast cells. [See, e.g., procedures described in published PCT application W086/00639 and European patent application EPA 123,289].

35 A method for producing high levels of a BMP-9 protein of the invention in mammalian cells may involve the construction of cells containing multiple copies of the heterologous BMP-9 gene. The heterologous gene is linked to an amplifiable marker, e.g. the dihydrofolate reductase (DHFR) gene for which cells containing increased gene copies can be selected for propagation in increasing concentrations of methotrexate (MTX) according to the procedures of Kaufman and Sharp, J. Mol. Biol., 159:601-629 (1982). This approach can be employed with a number of different cell types.

40 For example, a plasmid containing a DNA sequence for a BMP-9 of the invention in operative association with other plasmid sequences enabling expression thereof and the DHFR expression plasmid pAdA26SV(A)3 [Kaufman and Sharp, Mol. Cell. Biol., 2:1304 (1982)] can be co-introduced into DHFR-deficient CHO cells, DUKX-BII, by various methods including calcium phosphate coprecipitation and transfection, electroporation or protoplast fusion. DHFR expressing transformants are selected for growth in alpha media with dialyzed fetal calf serum, and subsequently selected for amplification by growth in increasing concentrations of MTX (e.g. sequential steps in 0.02, 0.2, 1.0 and 5uM MTX) as described in Kaufman et al., Mol Cell Biol., 5:1750 (1983). Transformants are cloned, and biologically active BMP-9 expression is monitored by the Rosen-modified Sampath - Reddi rat bone formation assay described above in Example III. BMP-9 expression should increase with increasing levels of MTX resistance. BMP-9 polypeptides are characterized using standard techniques known in the art such as pulse labeling with [35S] methionine or cysteine and polyacrylamide gel electrophoresis. Similar procedures can be followed to produce other related BMP-9 proteins.

A. BMP-9 Vector Construction

55 In order to produce human BMP-9 proteins of the invention DNA sequences encoding the mature region of the human BMP-9 protein may be joined to DNA sequences encoding the propeptide region of the murine BMP-9 protein. This murine/human hybrid DNA sequence is inserted into an appropriate expression vector and introduced into mammalian cells or other preferred eukaryotic or prokaryotic hosts by conventional genetic engineering techniques. The

construction of this murine/human BMP-9 containing expression plasmid is described below.

A derivative of the human BMP-9 sequence (SEQ ID NO:8) comprising the nucleotide sequence from nucleotide #105 to #470 is specifically amplified. The following oligonucleotides are utilized as primers to allow the amplification of nucleotides #105 to #470 of the human BMP-9 sequence (SEQ ID NO:8) from clone pGEM-111 described above.

#3 ATCGGGCCCCTTTTAGCCAGGCGGAAAAGGAG

#4 AGCGAATTCCCCGCAGGCAGATACTACCTG

This procedure generates the insertion of the nucleotide sequence ATCGGGCCCCT immediately preceding nucleotide #105 and the insertion of the nucleotide sequence GAATTCGCT immediately following nucleotide #470. The addition of these sequences results in the creation of an Apa I and EcoR I restriction endonuclease site at the respective ends of the specifically amplified DNA fragment. The resulting 374 bp Apa I/EcoR I fragment is subcloned into the plasmid vector pGEM-7Zf(+) (Promega catalog# p2251) which has been digested with Apa I and EcoR I. The resulting clone is designated pHMP9mex-1.

The following oligonucleotides are designed on the basis of murine BMP-9 sequences (SEQ ID NO:1) and are modified to facilitate the construction of the murine/human expression plasmid referred to above:

#5

GATTCGTCGACCACCATGTCCCCTGGGGCCTGGTCTAGATGGATACACAGCTGTGGGGCC

#6 CCACAGCTGTGTATCCATCTAGACCAGGCCCCAGGGGACATGGTGGTTCGACG

These oligonucleotides contain complimentary sequences which upon addition to each other facilitate the annealing (base pairing) of the two individual sequences, resulting in the formation of a double stranded synthetic DNA linker (designated LINK-1) in a manner indicated below:

	1	5	10	20	30	40	50	60
#5	G	A	T	T	C	G	T	C
	G	A	C	C	A	C	C	A
	T	G	T	C	C	C	T	G
	G	G	G	G	C	C	T	G
	G	G	T	C	T	A	G	A
	T	G	G	A	T	A	C	A
	C	A	G	C	T	G	T	G
	G	G	G	G	C	C	C	G
	A	C	C	C	G	A	C	A
	G	A	T	C	T	A	C	T
	A	C	T	A	T	G	T	G
	T	C	G	A	C	C		
#6								

This DNA linker (LINK-1) contains recognition sequences of restriction endonucleases needed to facilitate subsequent manipulations required to construct the murine/human expression plasmid, as well as sequences required for maximal expression of heterologous sequences in mammalian cell expression systems. More specifically (referring to the sequence numbering of oligonucleotide #5/LINK-1): nucleotides #1-#11 comprise recognition sequences for the restriction endonucleases BamH I and Sal I, nucleotides #11-#15 allow for maximal expression of heterologous sequences in mammalian cell expression systems, nucleotides #16-#31 correspond to nucleotides #610-#625 of the murine BMP-9 sequence (SEQ ID NO:1), nucleotides #32-#33 are inserted to facilitate efficient restriction digestion of two adjacent restriction endonuclease sites (EcoO109 I and Xba I), nucleotides #34-#60 correspond to nucleotides #1515-#1541 of the murine BMP-9 sequence (SEQ ID NO:1) except that nucleotide #58 of synthetic oligonucleotide #5 is a G rather than the A which appears at position #1539 of SEQ ID NO:1 (This nucleotide conversion results in the creation of an Apa I restriction endonuclease recognition sequence, without altering the amino acid sequence it is intended to encode, to facilitate further manipulations of the murine/human hybrid expression plasmid. LINK-1 (the double stranded product of the annealing of oligonucleotides #5 and #6) is subcloned into the plasmid vector pGEM-7Zf(+) which has been digested with the restriction endonucleases Apa I and BamH I. This results in a plasmid in which the sequences normally present between the Apa I and BamH I sites of the pGEM-7Zf(+) plasmid polylinker are replaced with the sequences of LINK-1 described above. The resulting plasmid clone is designated pBMP-9link.

pBMP-9link is digested with the restriction endonucleases BamH I and Xba I resulting in the removal nucleotides #1-#34 of LINK-1 (refer to the numbering of oligo #5). Clone ML14a, which contains an insert comprising the sequence

set forth in SEQ ID NO:1, is also digested with the restriction endonucleases BamH I and Xba I resulting in the removal of sequences comprising nucleotides #1-#1515 of SEQUENCE ID NO:1 (murine BMP-9). This BamH I/Xba I fragment of mouse BMP-9 is isolated from the remainder of the ML14a plasmid clone and subcloned into the BamH I/Xba I sites generated by the removal of the synthetic linker sequences described above. The resulting clone is designated p302.

The p302 clone is digested with the restriction endonuclease EcoO109 I resulting in the excision of nucleotides corresponding to nucleotides #621-#1515 of the murine BMP-9 sequence (SEQ ID NO:1) and nucleotides #35-#59 of LINK-1 (refer to numbering of oligonucleotide #5). It should be noted that the Apa I restriction site created in LINK-1 by the A to G conversion described above is a subset of the recognition sequence of EcoO109 I, therefore digestion of p302 with EcoO109 I cleaves at the Apa I site as well as the naturally occurring murine EcoO109 I (location #619-#625 of SEQ ID NO:1) resulting in the excision of a 920 bp EcoO109 I/EcoO109 I (Apa I) fragment comprising the sequences described above. This 920 EcoO109 I/EcoO109 I (Apa I) fragment is isolated from the remainder of the p302 plasmid clone and subcloned into clone pBMP-9link which has been similarly digested with EcoO109 I. It should be noted that the nucleotides GG (#32-#33 of oligonucleotide #5) originally designed to facilitate a more complete digestion of the two adjacent restriction sites EcoO109 I and Xba I of LINK-1, which is now a part of pBMP-9link (described above), results in the creation of Dcm methylation recognition sequence. The restriction nuclease EcoO109 I is sensitive to Dcm methylation and therefore cleavage of this sequence (nucleotides #25-#31 of oligonucleotide #5/LINK-1) by the restriction endonuclease EcoO109 I is prevented at this site. Therefore the plasmid clone pBMP-9link is cleaved at the Apa I site but not at the EcoO109 I site upon digestion with the restriction endonuclease EcoO109 I as described above, preventing the intended removal of the sequences between the EcoO109 I and Xba I site of LINK-1 (#32-#55 defined by the numbering of oligonucleotide #5). This results in the insertion of the 920 bp EcoO109 I/Apa I fragment at the EcoO109 I (Apa I) site of pBMP-9link. The resulting clone is designated p318.

Clone p318 is digested with the restriction endonucleases Sal I and Apa I, resulting in the excision of sequences comprising nucleotides #6-#56 of LINK-1 (refer to oligo #5 for location), nucleotides #621-#1515 of murine BMP-9 (SEQ ID NO:1), and nucleotides #35-#60 of LINK-1 (refer to oligo #5 for location). The resulting 972 bp Sal I/Apa I fragment described above is isolated from the remainder of the p318 plasmid clone and will be utilized in subsequent manipulations.

The clone phBMP9mex-1 (described above), which contains DNA sequences which encode the entire mature region and portions of the propeptide of the human BMP-9 protein, is digested with the restriction endonucleases Apa I and EcoR I. This results in the excision of a 374 bp fragment comprising nucleotides #105-#470 of the human BMP-9 sequence (SEQ ID NO:8) and the additional nucleotides of oligonucleotide primers #3 and #4 which contain the recognition sequences for the restriction endonucleases Apa I and EcoR I. This 374 bp Apa I/EcoR I fragment is combined with the 972 bp Sal I/Apa I fragment from p138 (isolation described above) and ligated to the mammalian cell expression plasmid pED6 (a derivative of pEMC2β1) which has been digested with Sal I and EcoR I. The resulting clone is designated p324.

The clone ML14a (murine BMP-9) is digested with EcoO109 I and Xba I to generate a fragment comprising nucleotides #621-#1515 of SEQ ID NO:1.

The following oligonucleotides are synthesized on an automated DNA synthesizer and combined such that their complementary sequences can base pair (anneal) with each other to generate a double stranded synthetic DNA linker designated LINK-2:

#7 TCGACCACCATGTCCCCTGG

#8 GCCCCAGGGGACATGGTGG

This double stranded synthetic DNA linker (LINK-2) anneals in such a way that it generates single stranded ends which are compatible to DNA fragments digested with Sal I (one end) or EcoO109 I (the other end) as indicated below:

#7 TCGACCACCATGTCCCCTGG
GGTGCTACAGGGGACCCCG #8

This LINK-2 synthetic DNA linker is ligated to the 895 bp EcoO109 I/Xba I fragment comprising nucleotides #621-#1515 of murine BMP-9 (SEQ ID NO:1) described above. This results in a 915 bp Sal I/Xba I fragment.

The clone p324 is digested with Sal I/Xba I to remove sequences comprising nucleotides #6-#56 of LINK-1 (refer to oligo #5 for location) and nucleotides #621-#1515 of murine BMP-9 (SEQ ID NO:1). The sequences comprising

nucleotides #35-#60 of LINK-1 (refer to oligo #5 for location) and the sequences comprising the 374 bp Apa I/EcoR I fragment (human BMP-9 sequences) derived from pHMP9mex-1 remain attached to the pED6 backbone. The 915 bp Sal I/Xba I fragment comprising LINK-2 sequences and nucleotides #621-#1515 of murine BMP-9 (SEQ ID NO:1) is ligated into the p324 clone from which the Sal I to Xba I sequences described above have been removed.

The resulting plasmid is designated BMP9fusion and comprises LINK-2, nucleotides #621-#1551 of murine BMP-9 (SEQ ID NO:1), nucleotides #35-#59 of LINK-1 (refer to the numbering of oligonucleotide #5), and the 374 bp Apa I/EcoR I fragment (human BMP-9) derived from clone pHMP9mex-1 (described above) inserted between the Sal I and EcoR I sites of the mammalian cell expression vector pED6.

BMP9 fusion is transfected into CHO cells using standard techniques known to those having ordinary skill in the art to create stable cell lines capable of expressing human BMP-9 protein. The cell lines are cultured under suitable culture conditions and the BMP-9 protein is isolated and purified from the culture medium.

EXAMPLE V

Biological Activity of Expressed BMP-9

To measure the biological activity of the expressed BMP-9 proteins obtained in Example IV above, the proteins are recovered from the cell culture and purified by isolating the BMP-9 proteins from other proteinaceous materials with which they are co-produced as well as from other contaminants. The purified protein may be assayed in accordance with the rat bone formation assay described in Example III.

Purification is carried out using standard techniques known to those skilled in the art. It is contemplated, as with other BMP proteins, that purification may include the use of Heparin sepharose.

Protein analysis is conducted using standard techniques such as SDS-PAGE acrylamide [U.K. Laemmli, *Nature* 227:680 (1970)] stained with silver [R.R. Oakley, et al. *Anal. Biochem.* 105:361 (1980)] and by immunoblot [H. Towbin, et al. *Proc. Natl. Acad. Sci. USA* 76:4350 (1979)]

The foregoing descriptions detail presently preferred embodiments of the present invention. Numerous modifications and variations in practice thereof are expected to occur to those skilled in the art upon consideration of these descriptions.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT: Wozney, John M. Celeste, Anthony

(ii) TITLE OF INVENTION: BMP-9 COMPOSITIONS

(iii) NUMBER OF SEQUENCES: 9

(iv) CORRESPONDENCE ADDRESS:

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(B) STREET: Legal Affairs - 87 CambridgePark Drive
(C) CITY: Cambridge
(D) STATE: MA
(E) COUNTRY: US
(F) ZIP: 02140

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.25

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER: US

(B) FILING DATE:
(C) CLASSIFICATION:

5 (viii) ATTORNEY/AGENT INFORMATION:

(A) NAME: Kapinos, Ellen J.
(B) REGISTRATION NUMBER: 32,245
(C) REFERENCE/DOCKET NUMBER: GI 5186A

10 (ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE: (617) 876-1170
(B) TELEFAX: (617) 876-5851

15 (2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

20 (A) LENGTH: 2447 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

25 (ii) MOLECULE TYPE: cDNA to mRNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

30 (vi) ORIGINAL SOURCE:

(A) ORGANISM: Mus musculus
(B) STRAIN: C57B46xCBA
(F) TISSUE TYPE: liver

35 (vii) IMMEDIATE SOURCE:

(A) LIBRARY: Mouse liver cDNA
(B) CLONE: ML14A

40 (viii) POSITION IN GENOME:

(C) UNITS: bp

45 (ix) FEATURE:

(A) NAME/KEY: mat_peptide
(B) LOCATION: 1564..1893

50 (ix) FEATURE:

(A) NAME/KEY: CDS
(B) LOCATION: 610..1896

55 (ix) FEATURE:

(A) NAME/KEY: mRNA
(B) LOCATION: 1..2447

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

5 CATTAAATAAA TATTAAGTAT TGGAATTAGT GAAATTGGAG TTCCTGTGG AAGGAACTGG 60
 GCAAGTGAGC TTTTITAGTTT GTGTGCGAAG CCTGTAATTA CGGCTCCAGC TCATAGTGGA 120
 ATGGCTATAC TTAGATTTAT GGATAGTTGG GTAGTAGGTG TAAATGTATG TGGTAAAAGG 180
 10 CCTAGGAGAT TTGTTGATCC AATAAATATG ATTACGGAAA CAATTATTAG GGTTCATGTT 240
 CGTCCTTTTG GTGTGTGGAT TAGCATTATT TGTTCGATAA TAAGTTTAAC TAGTCAGTGT 300
 TGGAAAGAAT GGAGACGGTT GTTGATTAGG CGTTTGTAGG ATGGGAATAG GATTGAAGGA 360
 AATATAATGA TGGCTACAAC GATTGGGAAT CCTATTATTG TTGGGGTAAT GAATGAGGCA 420
 15 AATAGATTTT CGTTCATTTT AATTCTCAAG GGGTTTTTAC TTTTATGTTT GTTAGTGATA 480
 TTGGTGAGTA GGCCAAGGGT TAATAGTGTA ATTGAATTAT AGTGAAATCA TATTACTAGA 540
 20 CCTGATGTTA GAAGGAGGGC TGAAAAGGCT CCTTCCCTCC CAGGACAAAA CCGGAGCAGG 600
 GCCACCCGG ATG TCC CCT GGG GCC TTC CGG GTG GCC CTG CTC CCG CTG 648
 Met Ser Pro Gly Ala Phe Arg Val Ala Leu Leu Pro Leu
 -318 -315 -310
 25 TTC CTG CTG GTC TGT GTC ACA CAG CAG AAG CCG CTG CAG AAC TGG GAA 696
 Phe Leu Leu Val Cys Val Thr Gln Gln Lys Pro Leu Gln Asn Trp Glu
 -305 -300 -295 -290
 CAA GCA TCC CCT GGG GAA AAT GCC CAC AGC TCC CTG GGA TTG TCT GGA 744
 Gln Ala Ser Pro Gly Glu Asn Ala His Ser Ser Leu Gly Leu Ser Gly
 30 -285 -280 -275
 GCT GGA GAG GAG GGT GTC TTT GAC CTG CAG ATG TTC CTG GAG AAC ATG 792
 Ala Gly Glu Glu Gly Val Phe Asp Leu Gln Met Phe Leu Glu Asn Met
 -270 -265 -260
 35 AAG GTG GAT TTC CTA CGC AGC CTT AAC CTC AGC GGC ATT CCC TCC CAG 840
 Lys Val Asp Phe Leu Arg Ser Leu Asn Leu Ser Gly Ile Pro Ser Gln

(2) INFORMATION FOR SEQ ID NO:9:

40

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 151 amino acids

(B) TYPE: amino acid

45

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

50

55

* Thr Arg Glu Cys Ser Arg Ser Cys Pro Arg Thr Ala Pro Gln Arg
 -41 -40 -35 -30
 5 Gln Val Arg Ala Val Thr Arg Arg Thr Arg Met Ala His Val Ala Ala
 -25 -20 -15 -10
 Gly Ser Thr Leu Ala Arg Arg Lys Arg Ser Ala Gly Ala Gly Ser His
 -5 1 5
 10 Cys Gln Lys Thr Ser Leu Arg Val Asn Phe Glu Asp Ile Gly Trp Asp
 10 15 20
 Ser Trp Ile Ile Ala Pro Lys Glu Tyr Glu Ala Tyr Glu Cys Lys Gly
 25 30 35
 15 Gly Cys Phe Phe Pro Leu Ala Asp Asp Val Thr Pro Thr Lys His Ala
 40 45 50 55
 Ile Val Gln Thr Leu Val His Leu Lys Phe Pro Thr Lys Val Gly Lys
 60 65 70
 20 Ala Cys Cys Val Pro Thr Lys Leu Ser Pro Ile Ser Val Leu Tyr Lys
 75 80 85
 Asp Asp Met Gly Val Pro Thr Leu Lys Tyr His Tyr Glu Gly Met Ser
 90 95 100
 25 Val Ala Glu Cys Gly Cys Arg
 105 110

30

(ix) FEATURE:

(A) NAME/KEY: exon

(B) LOCATION: 1..470

35

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1..456

40

(ix) FEATURE:

(A) NAME/KEY: mat_peptide

(B) LOCATION: 124..453

45

(ix) FEATURE:

(A) NAME/KEY: mRNA

(B) LOCATION: 1..470

50

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

55

	TGA ACA AGA GAG TGC TCA AGA AGC TGT CCA AGG ACG GCT CCA CAG AGG	48
	* Thr Arg Glu Cys Ser Arg Ser Cys Pro Arg Thr Ala Pro Gln Arg	
	-41 -40 -35 -30	
5	CAG GTG AGA GCA GTC ACG AGG AGG ACA CGG ATG GCG CAC GTG GCT GCG	96
	Gln Val Arg Ala Val Thr Arg Arg Thr Arg Met Ala His Val Ala Ala	
	-25 -20 -15 -10	
10	GGG TCG ACT TTA GCC AGG CGG AAA AGG AGC GCC GGG GCT GGC AGC CAC	144
	Gly Ser Thr Leu Ala Arg Arg Lys Arg Ser Ala Gly Ala Gly Ser His	
	-5 1 1 5	
	TGT CAA AAG ACC TCC CTG CGG GTA AAC TTC GAG GAC ATC GGC TGG GAC	192
	Cys Gln Lys Thr Ser Leu Arg Val Asn Phe Glu Asp Ile Gly Trp Asp	
	10 15 20	
15	AGC TGG ATC ATT GCA CCC AAG GAG TAT GAA GCC TAC GAG TGT AAG GGC	240
	Ser Trp Ile Ile Ala Pro Lys Glu Tyr Ala Tyr Glu Cys Lys Gly	
	25 30 35	
20	GGC TGC TTC TTC CCC TTG GCT GAC GAT GTG ACG CCG ACG AAA CAC GCT	288
	Gly Cys Phe Phe Pro Leu Ala Asp Asp Val Thr Pro Thr Lys His Ala	
	40 45 50 55	
	ATC GTG CAG ACC CTG GTG CAT CTC AAG TTC CCC ACA AAG GTG GGC AAG	336
	Ile Val Gln Thr Leu Val His Leu Lys Phe Pro Thr Lys Val Gly Lys	
	60 65 70	
25	GCC TGC TGT GTG CCC ACC AAA CTG AGC CCC ATC TCC GTC CTC TAC AAG	384
	Ala Cys Cys Val Pro Thr Lys Leu Ser Pro Ile Ser Val Leu Tyr Lys	
	75 80 85	
	GAT GAC ATG GGG GTG CCC ACC CTC AAG TAC CAT TAC GAG GGC ATG AGC	432
	Asp Asp Met Gly Val Pro Thr Leu Lys Tyr His Tyr Glu Gly Met Ser	
	90 95 100	
30	GTG GCA GAG TGT GGG TGC AGG TAGTATCTGC CTGCGGG	470
	Val Ala Glu Cys Gly Cys Arg	
	105 110	

35

CATGGGCAGC TCGAG

40 (2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

- 45 (A) LENGTH: 34 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

50 (ii) MOLECULE TYPE: cDNA to mRNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

55 CTGCAGGCCGA GCCTGAATTC CTCGAGCCAT CATG 34

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 68 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

CGAGGTTAAA AAACGTCTAG GCCCCCGAA CCACGGGGAC GTGGTTTTCC TTTGAAAAAC 60
 ACGATTGC 68

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 470 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(v) FRAGMENT TYPE: C-terminal

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Homo sapiens
 (H) CELL LINE: W138 (genomic DNA)

(vii) IMMEDIATE SOURCE:

- (A) LIBRARY: human genomic library
 (B) CLONE: lambda 111-1

(viii) POSITION IN GENOME:

- (C) UNITS: bp

Arg Ile Asn Ile Tyr Glu Val Met Lys Pro Pro Ala Glu Val Val Pro
 -115 -110 -105
 5 Gly His Leu Ile Thr Arg Leu Leu Asp Thr Arg Leu Val His His Asn
 -100 -95 -90 -85
 Val Thr Arg Trp Glu Thr Phe Asp Val Ser Pro Ala Val Leu Arg Trp
 -80 -75 -70
 10 Thr Arg Glu Lys Gln Pro Asn Tyr Gly Leu Ala Ile Glu Val Thr His
 -65 -60 -55
 Leu His Gln Thr Arg Thr His Gln Gly Gln His Val Arg Ile Ser Arg
 -50 -45 -40
 15 Ser Leu Pro Gln Gly Ser Gly Asn Trp Ala Gln Leu Arg Pro Leu Leu
 -35 -30 -25
 Val Thr Phe Gly His Asp Gly Arg Gly His Ala Leu Thr Arg Arg Arg
 -20 -15 -10 -5
 20 Arg Ala Lys Arg Ser Pro Lys His His Ser Gln Arg Ala Arg Lys Lys
 1 5 10
 Asn Lys Asn Cys Arg Arg His Ser Leu Tyr Val Asp Phe Ser Asp Val
 15 20 25
 Gly Trp Asn Asp Trp Ile Val Ala Pro Pro Gly Tyr Gln Ala Phe Tyr
 30 35 40
 30 Cys His Gly Asp Cys Pro Phe Pro Leu Ala Asp His Leu Asn Ser Thr
 45 50 55 60
 Asn His Ala Ile Val Gln Thr Leu Val Asn Ser Val Asn Ser Ser Ile
 65 70 75
 35 Pro Lys Ala Cys Cys Val Pro Thr Glu Leu Ser Ala Ile Ser Met Leu
 80 85 90
 Tyr Leu Asp Glu Tyr Asp Lys Val Val Leu Lys Asn Tyr Gln Glu Met
 95 100 105
 40 Val Val Glu Gly Cys Gly Cys Arg
 110 115

45 (2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- 50 (A) LENGTH: 15 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

55 (ii) MOLECULE TYPE: cDNA to mRNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

TGT GGG TGC CGC TGAGATCAGG CAGTCCTTGA GGATAGACAG ATATACACAC 1666
 Cys Gly Cys Arg
 115
 5 CACACACACA CACCACATAC ACCACACACA CACGTTCCCA TCCACTCACC CACACACTAC 1726
 ACAGACTGCT TCCTTATAGC TGGACTTTTA TTTAAAAAAA AAAAAAAAAA AATGGAAAAA 1786
 ATCCCTAAAC ATTCACCTTG ACCTTATTTA TGACTTTACG TGCAAATGTT TTGACCATAT 1846
 10 TGATCATATA TTTTGACAAA ATATATTTAT AACTACGTAT TAAAAGAAAA AAATAAAATG 1906
 AGTCATTATT TTAATAAAAA AAAAAAAACT CTAGAGTCGA CGGAATTC 1954

15 (2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

20 (A) LENGTH: 408 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Ile Pro Gly Asn Arg Met Leu Met Val Val Leu Leu Cys Gln Val
 -292 -290 -285 -280
 30 Leu Leu Gly Gly Ala Ser His Ala Ser Leu Ile Pro Glu Thr Gly Lys
 -275 -270 -265
 Lys Lys Val Ala Glu Ile Gln Gly His Ala Gly Gly Arg Arg Ser Gly
 -260 -255 -250 -245
 35 Gln Ser His Glu Leu Leu Arg Asp Phe Glu Ala Thr Leu Leu Gln Met
 -240 -235 -230
 Phe Gly Leu Arg Arg Arg Pro Gln Pro Ser Lys Ser Ala Val Ile Pro
 -225 -220 -215
 40 Asp Tyr Met Arg Asp Leu Tyr Arg Leu Gln Ser Gly Glu Glu Glu Glu
 -210 -205 -200
 Glu Gln Ile His Ser Thr Gly Leu Glu Tyr Pro Glu Arg Pro Ala Ser
 -195 -190 -185
 45 Arg Ala Asn Thr Val Arg Ser Phe His His Glu Glu His Leu Glu Asn
 -180 -175 -170 -165
 Ile Pro Gly Thr Ser Glu Asn Ser Ala Phe Arg Phe Leu Phe Asn Leu
 -160 -155 -150
 50 Ser Ser Ile Pro Glu Asn Glu Val Ile Ser Ser Ala Glu Leu Arg Leu
 -145 -140 -135
 Phe Arg Glu Gln Val Asp Gln Gly Pro Asp Trp Glu Arg Gly Phe His
 -130 -125 -120
 55

	GTG GAC CAG GGC CCT GAT TGG GAA AGG GGC TTC CAC CGT ATA AAC ATT	942
	Val Asp Gln Gly Pro Asp Trp Glu Arg Gly Phe His Arg Ile Asn Ile	
	-125 -120 -115	
5	TAT GAG GTT ATG AAG CCC CCA GCA GAA GTG GTG CCT GGG CAC CTC ATC	990
	Tyr Glu Val Met Lys Pro Pro Ala Glu Val Val Pro Gly His Leu Ile	
	-110 -105 -100	
10	ACA CGA CTA CTG GAC ACG AGA CTG GTC CAC CAC AAT GTG ACA CGG TGG	1038
	Thr Arg Leu Leu Asp Thr Arg Leu Val His His Asn Val Thr Arg Trp	
	-95 -90 -85	
	GAA ACT TTT GAT GTG AGC CCT GCG GTC CTT CGC TGG ACC CGG GAG AAG	1086
	Glu Thr Phe Asp Val Ser Pro Ala Val Leu Arg Trp Thr Arg Glu Lys	
	-80 -75 -70 -65	
15	CAG CCA AAC TAT GGG CTA GCC ATT GAG GTG ACT CAC CTC CAT CAG ACT	1134
	Gln Pro Asn Tyr Gly Leu Ala Ile Glu Val Thr His Leu His Gln Thr	
	-60 -55 -50	
20	CGG ACC CAC CAG GGC CAG CAT GTC AGG ATT AGC CGA TCG TTA CCT CAA	1182
	Arg Thr His Gln Gly Gln His Val Arg Ile Ser Arg Ser Leu Pro Gln	
	-45 -40 -35	
	GGG AGT GGG AAT TGG GCC CAG CTC CGG CCC CTC CTG GTC ACC TTT GGC	1230
	Gly Ser Gly Asn Trp Ala Gln Leu Arg Pro Leu Leu Val Thr Phe Gly	
	-30 -25 -20	
25	CAT GAT GGC CGG GGC CAT GCC TTG ACC CGA CGC CGG AGG GCC AAG CGT	1278
	His Asp Gly Arg Gly His Ala Leu Thr Arg Arg Arg Arg Ala Lys Arg	
	-15 -10 -5	
30	AGC CCT AAG CAT CAC TCA CAG CGG GCC AGG AAG AAG AAT AAG AAC TGC	1326
	Ser Pro Lys His His Ser Gln Arg Ala Arg Lys Lys Asn Lys Asn Cys	
	1 5 10 15	
	CGG CGC CAC TCG CTC TAT GTG GAC TTC AGC GAT GTG GGC TGG AAT GAC	1374
	Arg Arg His Ser Leu Tyr Val Asp Phe Ser Asp Val Gly Trp Asn Asp	
	20 25 30	
35	TGG ATT GTG GCC CCA CCA GGC TAC CAG GCC TTC TAC TGC CAT GGG GAC	1422
	Trp Ile Val Ala Pro Pro Gly Tyr Gln Ala Phe Tyr Cys His Gly Asp	
	35 40 45	
40	TGC CCC TTT CCA CTG GCT GAC CAC CTC AAC TCA ACC AAC CAT GCC ATT	1470
	Cys Pro Phe Pro Leu Ala Asp His Leu Asn Ser Thr Asn His Ala Ile	
	50 55 60	
	GTG CAG ACC CTG GTC AAT TCT GTC AAT TCC AGT ATC CCC AAA GCC TGT	1518
	Val Gln Thr Leu Val Asn Ser Val Asn Ser Ser Ile Pro Lys Ala Cys	
	65 70 75 80	
45	TGT GTG CCC ACT GAA CTG AGT GCC ATC TCC ATG CTG TAC CTG GAT GAG	1566
	Cys Val Pro Thr Glu Leu Ser Ala Ile Ser Met Leu Tyr Leu Asp Glu	
	85 90 95	
50	TAT GAT AAG GTG GTA CTG AAA AAT TAT CAG GAG ATG GTA GTA GAG GGA	1614
	Tyr Asp Lys Val Val Leu Lys Asn Tyr Gln Glu Met Val Val Glu Gly	
	100 105 110	

(B) LOCATION: 9..1934

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(xi) SAQUENCE DESCRIPTION: SEQ ID NO:3:

	CTCTAGAGGG CAGAGGAGGA GGGAGGGAGG GAAGGAGCGC GGAGCCCGGC CCGGAAGCTA	60
	GGTGAGTGTG GCATCCGAGC TGAGGGACGC GAGCCTGAGA CGCCGCTGCT GCTCCGGCTG	120
5	AGTATCTAGC TTGTCTCCCC GATGGGATTC CCGTCCAAGC TATCTCGAGC CTGCAGCGCC	180
	ACAGTCCCCG GCCCTCGCCC AGGTTCACTG CAACCGTTCA GAGGTCCCCA GGAGCTGCTG	240
	CTGGCGAGCC CGCTACTGCA GGGACCTATG GAGCCATTCC GTAGTGCCAT CCGAGCAAC	300
10	GCACTGCTGC AGCTTCCCTG AGCCTTTCCA GCAAGTTTGT TCAAGATTGG CTGTCAAGAA	360
	TCATGGACTG TTATTATATG CCTTGTTTTC TGTCAAGACA CC ATG ATT CCT GGT	414
	Met Ile Pro Gly	
	-292 -290	
15	AAC CGA ATG CTG ATG GTC GTT TTA TTA TGC CAA GTC CTG CTA GGA GGC	462
	Asn Arg Met Leu Met Val Val Leu Leu Cys Gln Val Leu Leu Gly Gly	
	-285 -280 -275	
	GCG AGC CAT GCT AGT TTG ATA CCT GAG ACG GGG AAG AAA AAA GTC GCC	510
20	Ala Ser His Ala Ser Leu Ile Pro Glu Thr Gly Lys Lys Lys Val Ala	
	-270 -265 -260	
	GAG ATT CAG GGC CAC GCG GGA GGA CGC CGC TCA GGG CAG AGC CAT GAG	558
	Glu Ile Gln Gly His Ala Gly Gly Arg Arg Ser Gly Gln Ser His Glu	
	-255 -250 -245	
25	CTC CTG CGG GAC TTC GAG GCG ACA CTT CTG CAG ATG TTT GGG CTG CGC	606
	Leu Leu Arg Asp Phe Glu Ala Thr Leu Leu Gln Met Phe Gly Leu Arg	
	-240 -235 -230 -225	
	CGC CGC CCG CAG CCT AGC AAG AGT GCC GTC ATT CCG GAC TAC ATG CGG	654
30	Arg Arg Pro Gln Pro Ser Lys Ser Ala Val Ile Pro Asp Tyr Met Arg	
	-220 -215 -210	
	GAT CTT TAC CGG CTT CAG TCT GGG GAG GAG GAG GAA GAG CAG ATC CAC	702
	Asp Leu Tyr Arg Leu Gln Ser Gly Glu Glu Glu Glu Glu Gln Ile His	
	-205 -200 -195	
35	AGC ACT GGT CTT GAG TAT CCT GAG CGC CCG GCC AGC CGG GCC AAC ACC	750
	Ser Thr Gly Leu Glu Tyr Pro Glu Arg Pro Ala Ser Arg Ala Asn Thr	
	-190 -185 -180	
	GTG AGG AGC TTC CAC CAC GAA GAA CAT CTG GAG AAC ATC CCA GGG ACC	798
40	Val Arg Ser Phe His His Glu Glu His Leu Glu Asn Ile Pro Gly Thr	
	-175 -170 -165	
	AGT GAA AAC TCT GCT TTT CGT TTC CTC TTT AAC CTC AGC AGC ATC CCT	846
	Ser Glu Asn Ser Ala Phe Arg Phe Leu Phe Asn Leu Ser Ser Ile Pro	
	-160 -155 -150 -145	
45	GAG AAC GAG GTG ATC TCC TCT GCA GAG CTT CGG CTC TTC CGG GAG CAG	894
	Glu Asn Glu Val Ile Ser Ser Ala Glu Leu Arg Leu Phe Arg Glu Gln	
	-140 -135 -130	

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				-10					-5					1			
	Gly	Ala	Ser ₅	Ser	His	Cys	Gln	Lys ₁₀	Thr	Ser	Leu	Arg	Val ₁₅	Asn	Phe	Glu	
5	Asp	Ile ₂₀	Gly	Trp	Asp	Ser	Trp ₂₅	Ile	Ile	Ala	Pro	Lys ₃₀	Glu	Tyr	Asp	Ala	
	Tyr ₃₅	Glu	Cys	Lys	Gly	Gly ₄₀	Cys	Phe	Phe	Pro	Leu ₄₅	Ala	Asp	Asp	Val	Thr ₅₀	
10	Pro	Thr	Lys	His	Ala	Ile	Val	Gln	Thr	Leu ₆₀	Val	His	Leu	Glu	Phe	Pro	
	Thr	Lys	Val	Gly ₇₀	Lys	Ala	Cys	Cys	Val ₇₅	Pro	Thr	Lys	Leu	Ser	Pro	Ile	
15	Ser	Ile	Leu ₈₅	Tyr	Lys	Asp	Asp	Met ₉₀	Gly	Val	Pro	Thr	Leu	Lys	Tyr	His	
	Tyr	Glu	Gly	Met	Ser	Val	Ala	Glu	Cys	Gly	Cys	Arg					
20		100					105					110					

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1954 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Homo sapiens
(G) CELL TYPE: Osteosarcoma Cell Line
(H) CELL LINE: U-20S

(vii) IMMEDIATE SOURCE:

- (A) LIBRARY: U20S cDNA in Lambda gtlO
(B) CLONE: Lambda U20S-3

(viii) POSITION IN GENOME:

- (C) UNITS: bp

(ix) **FEATURE:**

- (A) NAME/KEY: CDS
(B) LOCATION: 403..1629

(ix) FEATURE:

(A) NAME/KEY: mat_peptide

(B) LOCATION: 1279..1626

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(ix) FEATURE:

(A) NAME/KEY: mRNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

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Met Ser Pro Gly Ala Phe Arg Val Ala Leu Leu Pro Leu Phe Leu Leu
 -318 -315 -310 -305
 5 Val Cys Val Thr Gln Gln Lys Pro Leu Gln Asn Trp Glu Gln Ala Ser
 -300 -295 -290
 Pro Gly Glu Asn Ala His Ser Ser Leu Gly Leu Ser Gly Ala Gly Glu
 -285 -280 -275
 10 Glu Gly Val Phe Asp Leu Gln Met Phe Leu Glu Asn Met Lys Val Asp
 -270 -265 -260 -255
 Phe Leu Arg Ser Leu Asn Leu Ser Gly Ile Pro Ser Gln Asp Lys Thr
 -250 -245 -240
 15 Arg Ala Glu Pro Pro Gln Tyr Met Ile Asp Leu Tyr Asn Arg Tyr Thr
 -235 -230 -225
 Thr Asp Lys Ser Ser Thr Pro Ala Ser Asn Ile Val Arg Ser Phe Ser
 -220 -215 -210
 20 Val Glu Asp Ala Ile Ser Thr Ala Ala Thr Glu Asp Phe Pro Phe Gln
 -205 -200 -195
 Lys His Ile Leu Ile Phe Asn Ile Ser Ile Pro Arg His Glu Gln Ile
 -190 -185 -180 -175
 25 Thr Arg Ala Glu Leu Arg Leu Tyr Val Ser Cys Gln Asn Asp Val Asp
 -170 -165 -160
 Ser Thr His Gly Leu Glu Gly Ser Met Val Val Tyr Asp Val Leu Glu
 -155 -150 -145
 30 Asp Ser Glu Thr Trp Asp Gln Ala Thr Gly Thr Lys Thr Phe Leu Val
 -140 -135 -130
 Ser Gln Asp Ile Arg Asp Glu Gly Trp Glu Thr Leu Glu Val Ser Ser
 -125 -120 -115
 35 Ala Val Lys Arg Trp Val Arg Ala Asp Ser Thr Thr Asn Lys Asn Lys
 -110 -105 -100 -95
 Leu Glu Val Thr Val Gln Ser His Arg Glu Ser Cys Asp Thr Leu Asp
 -90 -85 -80
 40 Ile Ser Val Pro Pro Gly Ser Lys Asn Leu Pro Phe Phe Val Val Phe
 -75 -70 -65
 Ser Asn Asp Arg Ser Asn Gly Thr Lys Glu Thr Arg Leu Glu Leu Lys
 -60 -55 -50
 45 Glu Met Ile Gly His Glu Gln Glu Thr Met Leu Val Lys Thr Ala Lys
 -45 -40 -35
 Asn Ala Tyr Gln Val Ala Gly Glu Ser Gln Glu Glu Gly Leu Asp
 -30 -25 -20 -15
 50 Gly Tyr Thr Ala Val Gly Pro Leu Leu Ala Arg Arg Lys Arg Ser Thr
 55

	-15	-10	-5	
5	AGG AGC ACC GGA GCC AGC AGC CAC TGC CAG AAG ACT TCT CTC AGG GTG Arg Ser Thr Gly Ala Ser Ser His Cys Gln Lys Thr Ser Leu Arg Val 1 5 10 15	1608		
	AAC TTT GAG GAC ATC GGC TGG GAC AGC TGG ATC ATT GCA CCC AAG GAA Asn Phe Glu Asp Ile Gly Trp Asp Ser Trp Ile Ile Ala Pro Lys Glu 20 25 30	1656		
10	TAT GAC GCC TAT GAG TGT AAA GGG GGT TGC TTC TTC CCA TTG GCT GAT Tyr Asp Ala Tyr Glu Cys Lys Gly Cys Phe Phe Pro Leu Ala Asp 35 40 45	1704		
	GAC GTG ACA CCC ACC AAA CAT GCC ATC GTG CAG ACC CTG GTG CAT CTC Asp Val Thr Pro Thr Lys His Ala Ile Val Gln Thr Leu Val His Leu 50 55 60	1752		
	GAG TTC CCC ACA AAG GTG GGC AAA GCC TGC TGC GTT CCC ACC AAA CTG Glu Phe Pro Thr Lys Val Gly Lys Ala Cys Cys Val Pro Thr Lys Leu 65 70 75	1800		
20	AGT CCC ATC TCC ATC CTC TAC AAG GAT GAC ATG GGG GTG CCA ACC CTC Ser Pro Ile Ser Ile Leu Tyr Lys Asp Asp Met Gly Val Pro Thr Leu 80 85 90 95	1848		
25	AAG TAC CAC TAT GAG GGG ATG AGT GTG GCT GAG TGT GGG TGT AGG TAGTCCCTGC Lys Tyr His Tyr Glu Gly Met Ser Val Ala Glu Cys Gly Cys Arg 100 105 110	1904		
	AGCCACCCAG GGTGGGGATA CAGGACATGG AAGAGGTTCT GGTACGGTCC TGCATCCTCC	1963		
	TGCGCATGGT ATGCCTAAGT TGATCAGAAA CCATCCTTGA GAAGAAAAGG AGTTAGTTGC	2023		
30	CCTTCTTG TG TCTGGTGGGT CCCTCTGCTG AAGTGACAAT GACTGGGGTA TGCGGGCCTG	2083		
	TGGGCAGAGC AGGAGACCCT GGAAGGGTTA GTGGGTAGAA AGATGTCAAA AAGGAAGCTG	2143		
	TGGGTAGATG ACCTGCACTC CAGTGATTAG AAGTCCAGCC TTACCTGTGA GAGAGCTCCT	2203		
35	GGCATCTAAG AGAACTCTGC TTCCTCATCA TCCCCACCGA CTTGTTCTTC CTTGGGAGTG	2263		
	TGTCCTCAGG GAGAACAGCA TTGCTGTTCC TGTGCCTCAA GCTCCCAGCT GACTCTCCTG	2323		
	TGGCTCATAG GACTGAATGG GGTGAGGAAG AGCCTGATGC CCTCTGGCAA TCAGAGCCCG	2383		
40	AAGGACTTCA AAACATCTGG ACAACTCTCA TTGACTGATG CTCCAACATA ATTTTAAAA	2443		
	AGAG	2447		

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(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

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- (A) LENGTH: 428 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

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Claims

1. A DNA sequence encoding a protein having the biological activity of a BMP-9 protein of inducing the formation of

cartilage and/or bone which sequence is

- (a) the DNA sequence from nucleotides 124 to 453 of SEQ ID No. 8; or
 - (b) the DNA sequence from nucleotides 145 to 453 of SEQ ID No. 8; or
 - 5 (c) a DNA sequence which differs from the DNA sequence of (a) or (b) due to the degeneracies of the genetic code;
 - (d) an allelic variant of the sequence of (a) or (b); or
 - (e) a DNA sequence hybridizing under stringent conditions to the sequences of (a) or (b).
- 10 2. A recombinant DNA molecule containing a DNA sequence according to claim 1.
3. The recombinant DNA molecule according to claim 2 wherein said DNA sequence is under the control of regulatory elements allowing its expression in a desired host cell.
- 15 4. A host cell containing the recombinant DNA molecule according to claim 2 or 3.
5. The host cell according to claim 4 which is a bacterial cell, a yeast cell or a mammalian cell.
- 20 6. A method for the production of a protein having the biological activity of a BMP-9 protein comprising the cultivation of a host cell according to claim 4 or 5 under conditions appropriate for expression of said DNA sequence and recovering said protein from the culture.
7. A protein encoded by the DNA sequence of claim 1.
- 25 8. A protein produced by the method of claim 6.
9. A protein having the biological activity of a BMP-9 protein comprising one of the following amino acid sequences
 - (a) the amino acid sequence from amino acids No. 8 to 110 as set forth in Fig. 3 (SEQ ID No. 9); or
 - 30 (b) the amino acid sequence from amino acids No. 1 to 110 as set forth in Fig. 3 (SEQ ID No. 9).
10. A protein having the biological activity of a BMP-9 protein wherein said protein is a dimer wherein each subunit comprises at least the amino acid sequence from amino acids No. 8 to 110 of Fig. 3 (SEQ ID No. 9) or at least the amino acid sequence from amino acids No. 1 to 110 of Fig. 3 (SEQ ID No. 9).
- 35 11. A purified BMP-9 protein obtainable by the steps of
 - (a) culturing a cell transformed with a cDNA comprising the nucleotide sequence from nucleotides No. 124 to 453 as shown in Fig. 3 (SEQ ID No. 8); and
 - 40 (b) recovering and purifying from said culture medium a protein comprising the amino acid sequence from amino acids No. 1 to 110 as shown in Fig. 3 (SEQ ID No. 9).
12. A purified BMP-9 protein obtainable by the steps of
 - 45 (a) culturing a cell transformed with a cDNA comprising the nucleotide sequence from nucleotides No. 124 to 453 as shown in Fig. 3 (SEQ ID No. 8); and
 - (b) recovering from said culture medium a protein comprising an amino acid sequence from amino acids No. 8 to 110 as shown in Fig. 3 (SEQ ID No. 9).
- 50 13. A pharmaceutical composition comprising an effective amount of a protein according to any one of claims 7 to 12, optionally in conjunction with a pharmaceutically acceptable vehicle.
14. The composition of claim 13, further comprising a matrix for supporting said composition and providing a surface for bone and/or cartilage growth.
- 55 15. The composition of claim 14 wherein said matrix comprises a material which is hydroxyapatite, collagen, polylactic acid or tricalcium phosphate.

16. The pharmaceutical composition of any one of claims 13 to 15 for wound healing, tissue repair, inducing bone growth or inducing cartilage growth.
17. Use of a protein according to any one of claims 7 to 12 for preparing a pharmaceutical composition for inducing bone formation, cartilage formation, treatment of wounds or tissue repair.
18. A method for the preparation of a DNA sequence encoding a protein having the biological activity of a BMP-9 protein of inducing the formation of cartilage and/or bone which sequence is
- (a) the DNA sequence from nucleotides 124 to 453 of SEQ ID No. 8; or
 - (b) the DNA sequence from nucleotides 145 to 453 of SEQ ID No. 8; or
 - (c) a DNA sequence which differs from the DNA sequence of (a) or (b) due to degeneracies of the genetic code;
 - (d) an allelic variant of the sequence of (a) or (b); or
 - (e) a DNA sequence hybridizing under stringent conditions to the sequences of (a) or (b),
- said method comprising the following steps:
- (i) plating a human genomic library and preparing duplicate nitrocellulose replicas;
 - (ii) hybridizing one set of the duplicate nitrocellulose replicas with the labeled oligonucleotide
- #1: CTATGAGTGTAAGGGGGTTGCTTCTTCCCATTGGCTGAT
- and the other set with the labeled oligonucleotide
- #2: GTGCCAACCTCAAGTACCACTATGAGGGGATGAGTGTGG;
- (iii) isolating those clones which hybridize to both oligonucleotides and determining the sequence of their inserts.
19. A process for the manufacture of a composition according to claim 13, characterized in the use of the protein of any one of claims 7 to 12 as an essential constituent of said composition.

Patentansprüche

1. DNA-Sequenz, die ein Protein mit der biologischen Aktivität der Induktion der Bildung von Knorpel und/oder Knochen eines BMP-9-Proteins codiert, wobei die Sequenz ist
- (a) die DNA-Sequenz von Nucleotid 124 bis 453 von SEQ ID No. 8; oder
 - (b) die DNA-Sequenz von Nucleotid 145 bis 453 von SEQ ID No. 8; oder
 - (c) eine DNA-Sequenz, die sich von der DNA-Sequenz nach (a) oder (b) aufgrund der Degeneration des genetischen Codes unterscheidet; oder
 - (d) eine allelische Variante der Sequenz nach (a) oder (b); oder
 - (e) eine DNA-Sequenz, die unter stringenten Bedingungen mit den Sequenzen nach (a) oder (b) hybridisiert.
2. Rekombinantes DNA-Molekül, das eine DNA-Sequenz nach Anspruch 1 enthält.
3. Rekombinantes DNA-Molekül nach Anspruch 2, wobei die DNA-Sequenz unter der Kontrolle von regulatorischen Elementen steht, die ihre Expression in einer gewünschten Wirtszelle erlauben.
4. Wirtszelle, die das rekombinante DNA-Molekül nach Anspruch 2 oder 3 enthält.
5. Wirtszelle nach Anspruch 4, die eine Bakterienzelle, eine Hefezelle oder eine Säugerzelle ist.

6. Verfahren zur Herstellung eines Proteins mit der biologischen Aktivität eines BMP-9-Proteins, umfassend die Züchtung einer Wirtszelle nach Anspruch 4 oder 5 unter Bedingungen, die für die Expression der DNA-Sequenz geeignet sind, und die Gewinnung des Proteins aus der Kultur.
- 5 7. Protein, das von der DNA-Sequenz nach Anspruch 1 codiert wird.
8. Protein, das durch das Verfahren nach Anspruch 6 hergestellt wird.
9. Protein mit der biologischen Aktivität eines BMP-9-Proteins, das eine der folgenden Aminosäuresequenzen umfaßt
10 (a) die Aminosäuresequenz von Aminosäure Nr. 8 bis 110, die in Fig. 3 (SEQ ID No. 9) dargestellt ist; oder
(b) die Aminosäuresequenz von Aminosäure Nr. 1 bis 110, die in Fig. 3 (SEQ ID No. 9) dargestellt ist.
10. Protein mit der biologischen Aktivität eines BMP-9-Proteins, wobei das Protein ein Dimer ist, in dem jede Unter-
15 einheit mindestens die Aminosäuresequenz von Aminosäure Nr. 8 bis 110 von Fig. 3 (SEQ ID No. 9) oder mindestens die Aminosäuresequenz von Aminosäure Nr. 1 bis 110 von Fig. 3 (SEQ ID No. 9) umfaßt.
11. Gereinigtes BMP-9-Protein, erhältlich durch die Schritte
20 (a) Züchtung einer Zelle, die mit einer cDNA transformiert ist, die die Nucleotidsequenz von Nucleotid Nr. 124 bis 453 umfaßt, die in Fig. 3 (SEQ ID No. 8) gezeigt ist; und
(b) Gewinnung und Reinigung eines Proteins, das die Aminosäuresequenz von Aminosäure Nr. 1 bis 110 umfaßt, die in Fig. 3 (SEQ ID No. 9) gezeigt ist, aus dem Kulturmedium.
- 25 12. Gereinigtes BMP-9-Protein, erhältlich durch die Schritte
(a) Züchtung einer Zelle, die mit einer cDNA transformiert ist, die die Nucleotidsequenz von Nucleotid Nr. 124 bis 453 umfaßt, die in Fig. 3 (SEQ ID No. 8) gezeigt ist; und
(b) Gewinnung eines Proteins, das die Aminosäuresequenz von Aminosäure Nr. 8 bis 110 umfaßt, die in Fig.
30 3 (SEQ ID No. 9) gezeigt ist, aus dem Kulturmedium.
13. Arzneimittel, das eine wirksame Menge eines Proteins nach einem der Ansprüche 7 bis 12 gegebenenfalls in Verbindung mit einem pharmazeutisch verträglichen Träger, umfaßt.
- 35 14. Arzneimittel nach Anspruch 13, das weiter eine Matrix als Träger des Arzneimittels umfaßt und eine Oberfläche für Knochen- und/oder Knorpelwachstum bereitstellt.
15. Arzneimittel nach Anspruch 14, wobei die Matrix ein Material umfaßt, das Hydroxyapatit, Collagen, Polymilchsäure oder Tricalciumphosphat ist.
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16. Arzneimittel nach einem der Ansprüche 13 bis 15 zur Wundheilung, Gewebewiederherstellung, Induktion des Knochenwachstums oder Induktion des Knorpelwachstums.
17. Verwendung eines Proteins nach einem der Ansprüche 7 bis 12 zur Herstellung eines Arzneimittels zur Induktion der Knochenbildung oder der Knorpelbildung, zur Behandlung von Wunden oder zur Gewebewiederherstellung.
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18. Verfahren zur Herstellung einer DNA-Sequenz, die ein Protein mit der biologischen Aktivität der Induktion der Bildung von Knorpel und/oder Knochen eines BMP-9-Proteins codiert, wobei die Sequenz ist
50 (a) die DNA-Sequenz von Nucleotid 124 bis 453 von SEQ ID No. 8; oder
(b) die DNA-Sequenz von Nucleotid 145 bis 453 von SEQ ID No. 8; oder
(c) eine DNA-Sequenz, die sich von der DNA-Sequenz nach (a) oder (b) aufgrund der Degeneration des genetischen Codes unterscheidet; oder
(d) eine allelische Variante der Sequenz nach (a) oder (b); oder
55 (e) eine DNA-Sequenz, die unter stringenten Bedingungen mit den Sequenzen nach (a) oder (b) hybridisiert,

wobei das Verfahren die folgenden Schritte umfaßt:

- (i) Plattierung einer menschlichen genomischen Genbank und Herstellung von Nitrocellulose-Zweifachreplikas;
- (ii) Hybridisierung eines Satzes der Nitrocellulose-Zweifachreplikas mit dem markierten Oligonucleotid

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#1: CTATGAGTGTAAAGGGGGTTGCTTCTTCCCATTGGC.TGAT

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und des anderen Satzes mit dem markierten Oligonucleotid

#2: GTGCCAACCCTCAAGTACCACTATGAGGGGATGAGTGTGG;

15

und

(iii) Isolierung derjenigen Clone, die mit beiden Oligonucleotiden hybridisieren, und Bestimmung der Sequenz ihrer Insertionen.

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19. Verfahren zur Herstellung eines Mittels nach Anspruch 13, dadurch gekennzeichnet, daß man ein Protein nach einem der Ansprüche 7 bis 12 als wesentlichen Bestandteil des Mittels verwendet.

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Revendications

1. Séquence d'ADN codant pour une protéine ayant l'activité biologique d'une protéine BMP-9 d'induire la formation de cartilage et d'os, laquelle séquence est

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- (a) la séquence d'ADN des nucléotides 124 à 453 de SEQ ID No. 8; ou
- (b) la séquence d'ADN des nucléotides 145 à 453 de SEQ ID No. 8; ou
- (c) une séquence d'ADN qui diffère de la séquence d'ADN de (a) ou (b) due aux dégénérescences du code génétique;
- (d) une variante allèle de la séquence de (a) ou (b); ou
- (e) une séquence d'ADN s'hybridant sous des conditions rigoureuses en les séquences de (a) ou (b).

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2. Molécule d'ADN recombinant contenant une séquence d'ADN suivant la revendication 1.

3. Molécule d'ADN recombinant suivant la revendication 2, dans laquelle la séquence d'ADN est sous le contrôle d'éléments régulateurs permettant son expression dans une cellule hôte désirée.

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4. Cellule hôte contenant la molécule d'ADN recombinant suivant l'une ou l'autre des revendications 2 et 3.

5. Cellule hôte suivant la revendication 4, qui est une cellule bactérienne, une cellule de levure ou une cellule mam-mifère.

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6. Procédé de production d'une protéine ayant l'activité biologique d'une protéine BMP-9, comprenant la culture d'une cellule hôte suivant l'une ou l'autre des revendications 4 et 5 sous des conditions appropriées pour l'expression de la séquence d'ADN précitée et la récupération de ladite protéine de la culture.

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7. Protéine codée par la séquence d'ADN de la revendication 1.

8. Protéine produite par le procédé de la revendication 6.

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9. Protéine ayant l'activité biologique d'une protéine BMP-9 comprenant une des séquences d'acides aminés suivantes :

- (a) la séquence d'acides aminés allant des acides aminés n° 8 à 110 telle que représentée à la figure 3 (SEQ

ID No. 9); ou

(b) la séquence d'acides aminés allant des acides aminés n° 1 à 110 telle que représentée à la figure 3 (SEQ ID No. 9).

5 10. Protéine ayant l'activité biologique d'une protéine BMP-9, dans laquelle ladite protéine est un dimère dans lequel chaque sous-unité comprend au moins la séquence d'acides aminés allant des acides aminés n° 8 à 110 de la figure 3 (SEQ ID No. 9) ou au moins la séquence d'acides aminés allant des acides aminés n° 1 à 110 de la figure 3 (SEQ ID No. 9).

10 11. Protéine BMP-9 purifiée obtainable par les étapes suivantes :

(a) la culture d'une cellule transformée avec un ADNc comprenant la séquence nucléotidique allant des nucléotides n° 124 à n° 453 telle que représentée à la figure 3 (SEQ ID No. 8); et

15 (b) la récupération et la purification dudit milieu de culture d'une protéine comprenant la séquence d'acides aminés allant des acides aminés n° 1 à 110 telle que représentée à la figure 3 (SEQ ID No. 9).

12. Protéine BMP-9 purifiée obtainable par les étapes suivantes :

20 (a) la culture d'une cellule transformée avec un ADNc comprenant la séquence nucléotidique allant des nucléotides n° 124 à n° 453 telle que représentée à la figure 3 (SEQ ID No. 8); et

(b) la récupération dudit milieu de culture d'une protéine comprenant une séquence d'acides aminés allant des acides aminés n° 8 à 110 telle que représentée à la figure 3 (SEQ ID No. 9).

25 13. Composition pharmaceutique comprenant une quantité efficace d'une protéine suivant l'une quelconque des revendications 7 à 12, éventuellement conjointement à un véhicule pharmaceutiquement acceptable.

14. Composition suivant la revendication 13, comprenant de plus une matrice pour supporter ladite composition et former une surface pour la croissance d'os et/ou de cartilage.

30 15. Composition suivant la revendication 14, dans laquelle ladite matrice comprend une matière qui est une hydroxyapatite, du collagène, de l'acide polylactique ou du phosphate tricalcique.

16. Composition pharmaceutique suivant l'une quelconque des revendications 13 à 15, pour cicatriser les blessures, réparer les tissus, induire une croissance osseuse ou induire la croissance de cartilage.

35 17. Utilisation d'une protéine suivant l'une quelconque des revendications 7 à 12 pour préparer une composition pharmaceutique pour induire une formation osseuse, la formation de cartilage, le traitement de blessures ou la réparation de tissus.

40 18. Procédé de préparation d'une séquence d'ADN codant pour une protéine ayant l'activité biologique d'une protéine BMP-9 d'induire la formation de cartilage et/ou d'os, laquelle séquence est

(a) la séquence d'ADN des nucléotides 124 à 453 de SEQ ID No. 8; ou

(b) la séquence d'ADN des nucléotides 145 à 453 de SEQ ID No. 8; ou

45 (c) une séquence d'ADN qui diffère de la séquence d'ADN de (a) ou (b) due aux dégénérescences du code génétique;

(d) une variante allèle de la séquence de (a) ou (b); ou

(e) une séquence d'ADN s'hybridant sous des conditions rigoureuses en les séquences de (a) ou (b).

50 ledit procédé comprenant les étapes suivantes :

(i) l'étalement d'une bibliothèque génomique humaine et la préparation de répliques de nitrocellulose dupli-

quées;

55 (ii) l'hybridation d'une série des répliques de nitrocellulose dupliquées avec l'oligonucléotide marqué

n° 1 : CTATGAGTGTAAGGGGGTTGCTTCTTCCCATTGGCTGAT

et l'autre série avec l'oligonucléotide marqué

n° 2 : GTGCCAACCCTCAAGTACCACTATGAGGGGATGAGTGTGG; et

5

(iii) l'isolement de ces clones qui s'hybrident aux deux oligonucléotides et la détermination de la séquence de leurs inserts.

10 19. Procédé de fabrication d'une composition suivant la revendication 13, caractérisée par l'utilisation de la protéine suivant l'une quelconque des revendications 7 à 12 comme constituant essentiel de ladite composition.

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Figure 1A

10 20 30 40 50 60 70
 CATTAAATAA TATTAAGTAT TGGAATTAGT GAAATTGGAG TTCCTTGTGG AAGGAAGTGG GCAAGTGAGC
 80 90 100 110 120 130 140
 TTTTtagTTT GTGTCGGAAG CCTGTAATTA CGGCTCCAGC TCATAGTGGa ATGGCTATAC TTAGATTTAT
 150 160 170 180 190 200 210
 GGATAGTTGG GTAGTAGGTG TAAATGTATG TGGTAAAGG CCTAGGAGAT TTGTTGATCC AATAAATATG
 220 230 240 250 260 270 280
 ATTAGGGAAA CAATTATTAG GGTTCATGTT CGTCCTTTTG GTGTGTGGAT TAGCATTATT TGTTTGATAA
 290 300 310 320 330 340 350
 TAAGTTTAAC TAGTCAGTGT TGGAAAGAAT GGAGACGGTT GTTGATTAGG CGTTTTGAGG ATGGGAATAG
 360 370 380 390 400 410 420
 GATTGAAGGA AATATAATGA TGGCTACAAC GATTGGGAAT CCTATTATTG TTGGGGTAAT GAATGAGGCA
 430 440 450 460 470 480 490
 AATAGATTTT CGTTCATTTT AATTCTCAAG GGGTTTTTAC TTTTATGTTT GTTAGTGATA TTGGTGAGTA
 500 510 520 530 540 550 560
 GGCCAAGGGT TAATAGTGTA ATTGAATTAT AGTGAATCA TATTACTAGA CCTGATGTTA GAAGGAGGGC
 570 580 590 600 609 618
 TGAAAAGGCT CCTTCCCTCC CAGGACAAAA CCGGAGCAGG GCCACCCGG [>]ATG TCC CCT GGG
 M S P G
 627 636 645 654 663 672
 GCC TTC CGG GTG GCC CTG CTC CCG CTG TTC CTG CTG GTC TGT GTC ACA CAG CAG
 A F R V A L L P L F L L V C V T Q Q
 681 690 699 708 717 726
 AAG CCG CTG CAG AAC TGG GAA CAA GCA TCC CCT GGG GAA AAT GCC CAC AGC TCC
 K P L Q N W E Q A S P G E N A H S S
 735 744 753 762 771 780
 CTG GGA TTG TCT GGA GCT GGA GAG GAG GGT GTC TTT GAC CTG CAG ATG TTC CTG
 L G L S G A G E E G V F D L Q M F L
 789 798 807 816 825 834
 GAG AAC ATG AAG GTG GAT TTC CTA CGC AGC CTT AAC CTC AGC GGC ATT CCC TCC
 E N M K V D F L R S L N L S G I P S

Figure 1B

843	852	861	870	879	888
CAG GAC AAA ACC AGA GCG GAG CCA CCC CAG TAC ATG ATC GAC TTG TAC AAC AGA					
Q D K T R A E P P Q Y M I D L Y N R					
897	906	915	924	933	942
TAC ACA ACG GAC AAA TCG TCT ACG CCT GCC TCC AAC ATC GTG CGG AGC TTC AGC					
Y T T D K S S T P A S N I V R S P S					
951	960	969	978	987	996
GTG GAA GAT GCT ATA TCG ACA GCT GCC ACG GAG GAC TTC CCC TTT CAG AAG CAC					
V E D A I S T A A T E D F P F Q K H					
1005	1014	1023	1032	1041	1050
ATC CTG ATC TTC AAC ATC TCC ATC CCG AGG CAC GAG CAG ATC ACC AGG GCT GAG					
I L I F N I S I P R H E Q I T R A E					
1059	1068	1077	1086	1095	1104
CTC CGA CTC TAT GTC TCC TGC CAA AAT GAT GTG GAC TCC ACT CAT GGG CTG GAA					
L R L Y V S C Q N D V D S T H G L E					
1113	1122	1131	1140	1149	1158
GGA AGC ATG GTC GTT TAT GAT GTT CTG GAG GAC AGT GAG ACT TGG GAC CAG GCC					
G S M V V Y D V L E D S E T W D Q A					
1167	1176	1185	1194	1203	1212
ACG GGG ACC AAG ACC TTC TTG GTA TCC CAG GAC ATT CGG GAC GAA GGA TGG GAG					
T G T K T F L V S Q D I R D E G W E					
1221	1230	1239	1248	1257	1266
ACT TTA GAA GTA TCG AGT GCC GTG AAG CGG TGG GTC AGG GCA GAC TCC ACA ACA					
T L E V S S A V K R W V R A D S T T					
1275	1284	1293	1302	1311	1320
AAC AAA AAT AAG CTC GAG GTG ACA GTG CAG AGC CAC AGG GAG AGC TGT GAC ACA					
N K N K L E V T V Q S H R E S C D T					
1329	1338	1347	1356	1365	1374
CTG GAC ATC AGT GTC CCT CCA GGT TCC AAA AAC CTG CCC TTC TTT GTT GTC TTC					
L D I S V P P G S K N L P F F V V F					

Figure 1C

1383	1392	1401	1410	1419	1428
TCC AAT GAC CGC AGC AAT GGG ACC AAG GAG ACC AGA CTG GAG CTG AAG GAG ATC					
S N D R S N G T K E T R L E L K E M					
1437	1446	1455	1464	1473	1482
ATC GGC CAT GAG CAG GAG ACC ATG CTT GTG AAG ACA GCC AAA AAT GCT TAC CAG					
I G H E Q E T M L V K T A K N A Y Q					
1491	1500	1509	1518	1527	1536
GTG GCA GGT GAG AGC CAA GAG GAG GAG GGT CTA GAT GGA TAC ACA GCT GTG GGA					
V A G E S Q E E E G L D G Y T A V G					
1545	1554	1563	1572	1581	1590
CCA CTT TTA GCT AGA AGG AAG AGG AGC ACC GGA GCC AGC AGC CAC TGC CAG AAG					
P L L A R R K R S T G A S S H C Q K					
1599	1608	1617	1626	1635	1644
ACT TCT CTC AGG GTG AAC TTT GAG GAC ATC GGC TGG GAC AGC TGG ATC ATT GCA					
T S L R V N F E D I G W D S W I I A					
1653	1662	1671	1680	1689	1698
CCC AAG GAA TAT GAC GCC TAT GAG TGT AAA GGG GGT TGC TTC TTC CCA TTG GCT					
P K E Y D A Y E C K G G C F F P L A					
1707	1716	1725	1734	1743	1752
GAT GAC GTG ACA CCC ACC AAA CAT GCC ATC GTG CAG ACC CTG GTG CAT CTC GAG					
D D V T P T K H A I V Q T L V H L E					
1761	1770	1779	1788	1797	1806
TTC CCC ACA AAG GTG GGC AAA GCC TGC TGC GTT CCC ACC AAA CTG AGT CCC ATC					
F P C K V G K A C C V P T K L S P I					
1815	1824	1833	1842	1851	1860
TCC ATC CTC TAC AAG GAT GAC ATG GGG GTG CCA ACC CTC AAG TAC CAC TAT GAG					
S I L Y K D D M G V P T L K Y H Y E					
1869	1878	1887	1903	1913	1923
GGG ATG AGT GTG GCT GAG TGT GGG TGT AGG TAGTCCCTGC AGCCACCCAG GGTGGGGATA					
G M S T A E C G C R					

(428)

Figure 1D

1933	1943	1953	1963	1973	1983	1993
CAGGACATGG	AAGAGGTTCT	GGTACGGTCC	TGCATCCTCC	TGCGCATGGT	ATGCCTAAGT	TGATCAGAAA
2003	2013	2023	2033	2043	2053	2063
CCATCCTTGA	GAAGAAAAGG	AGTTAGTTGC	CCTTCTTGTC	TCTGGTGGGT	CCCTCTGCTG	AAGTGACAAT
2073	2083	2093	2103	2113	2123	2133
GACTGGGGTA	TGCGGGCCTG	TGGGCAGAGC	AGGAGACCCT	GGAAGGGTTA	GTGGGTAGAA	AGATGTCAAA
2143	2153	2163	2173	2183	2193	2203
AAGGAAGCTG	TGGGTAGATG	ACCTGCACTC	CAGTGATTAG	AAGTCCAGCC	TTACCTGTGA	GAGAGCTCCT
2213	2223	2233	2243	2253	2263	2273
GGCATCTAAG	AGAACTCTGC	TTCCTCATCA	TCCCCACCGA	CTTGTTCTTC	CTTGGGAGTG	TGTCCTCAGG
2283	2293	2303	2313	2323	2333	2343
GAGAACAGCA	TTGCTGTTCC	TGTGCCTCAA	GCTCCCAGCT	GACTCTCCTG	TGGCTCATAG	GACTGAATGG
2353	2363	2373	2383	2393	2403	2413
GGTGAGGAAG	AGCCTGATGC	CCTCTGGCAA	TCAGAGCCCG	AAGGACTTCA	AAACATCTGG	ACAACTCTCA
2423	2433	2443				
TTGACTGATG	CTCCAACATA	ATTTTAAAA	AGAG			

Figure 2

10 20 30 40 50 60 70
 CTCTAGAGGG CAGAGGAGGA GGGAGGGAGG GAAGGAGCGC GGAGCCCGGC CCGGAAGCTA GGTGAGTGTG
 80 90 100 110 120 130 140
 GCATCCGAGC TGAGGGACGC GAGCCTGAGA CGCCGCTGCT GCTCCGGCTG AGTATCTAGC TTGTCTCCCC
 150 160 170 180 190 200 210
 GATGGGATTG CCGTCCAAGC TATCTCGAGC CTGCAGCGCC ACAGTCCCCG GCCCTCGCCC AGGTTCACTG
 220 230 240 250 260 270 280
 CAACCGTTCA GAGGTCCCCA GGAGCTGCTG CTGGCGAGCC CGCTACTGCA GGGACCTATG GAGCCATTCC
 290 300 310 320 330 340 350
 GTAGTGCCAT CCCGAGCAAC GCACTGCTGC AGCTTCCCTG AGCCTTTCCA GCAAGTTTGT TCAAGATTGG
 360 370 380 390 400 (1)
 CTGTCAAGAA TCATGGACTG TTATTATATG CCTTGTTTTC TGTCAAGACA CC ATG ATT CCT
 MET Ile Pro
 417 432 447 462
 GGT AAC CGA ATG CTG ATG GTC GTT TTA TTA TGC CAA GTC CTG CTA GGA GGC GCG
 Gly Asn Arg MET Leu MET Val Val Leu Leu Cys Gln Val Leu Leu Gly Gly Ala
 477 492 507
 AGC CAT GCT AGT TTG ATA CCT GAG ACG GGG AAG AAA AAA GTC GCC GAG ATT CAG
 Ser His Ala Ser Leu Ile Pro Glu Thr Gly Lys Lys Lys Val Ala Glu Ile Gln
 522 537 552 567
 GGC CAC GCG GGA GGA CGC CGC TCA GGG CAG AGC CAT GAG CTC CTG CGG GAC TTC
 Gly His Ala Gly Gly Arg Arg Ser Gly Gln Ser His Glu Leu Leu Arg Asp Phe
 582 597 612 627
 GAG GCG ACA CTT CTG CAG ATG TTT GGG CTG CGC CGC CGC CCG CAG CCT AGC AAG
 Glu Ala Thr Leu Leu Gln MET Phe Gly Leu Arg Arg Arg Pro Gln Pro Ser Lys
 642 657 672
 AGT GCC GTC ATT CCG GAC TAC ATG CGG GAT CTT TAC CGG CTT CAG TCT GGG GAG
 Ser Ala Val Ile Pro Asp Tyr MET Arg Asp Leu Tyr Arg Leu Gln Ser Gly Glu
 687 702 717 732
 GAG GAG GAA GAG CAG ATC CAC AGC ACT GGT CTT GAG TAT CCT GAG CGC CCG GCC
 Glu Glu Glu Glu Gln Ile His Ser Thr Gly Leu Glu Tyr Pro Glu Arg Pro Ala
 747 762 777
 AGC CGG GCC AAC ACC GTG AGG AGC TTC CAC CAC GAA GAA CAT CTG GAG AAC ATC
 Ser Arg Ala Asn Thr Val Arg Ser Phe His His Glu Glu His Leu Glu Asn Ile
 792 807 822 837
 CCA GGG ACC AGT GAA AAC TCT GCT TTT CGT TTC CTC TTT AAC CTC AGC AGC ATC
 Pro Gly Thr Ser Glu Asn Ser Ala Phe Arg Phe Leu Phe Asn Leu Ser Ser Ile

Figure 2A

852 867 882 897
 CCT GAG AAC GAG GTG ATC TCC TCT GCA GAG CTT CGG CTC TTC CGG GAG CAG GTG
 Pro Glu Asn Glu Val Ile Ser Ser Ala Glu Leu Arg Leu Phe Arg Glu Gln Val

912 927 942
 GAC CAG GGC CCT GAT TGG GAA AGG GGC TTC CAC CGT ATA AAC ATT TAT GAG GTT
 Asp Gln Gly Pro Asp Trp Glu Arg Gly Phe His Arg Ile Asn Ile Tyr Glu Val

957 972 987 1002
 ATG AAG CCC CCA GCA GAA GTG GTG CCT GGG CAC CTC ATC ACA CGA CTA CTG GAC
 MET Lys Pro Pro Ala Glu Val Val Pro Gly His Leu Ile Thr Arg Leu Leu Asp

1017 1032 1047
 ACG AGA CTG GTC CAC CAC AAT GTG ACA CGG TGG GAA ACT TTT GAT GTG AGC CCT
 Thr Arg Leu Val His His Asn Val Thr Arg Trp Glu Thr Phe Asp Val Ser Pro

1062 1077 1092 1107
 GCG GTC CTT CGC TGG ACC CGG GAG AAG CAG CCA AAC TAT GGG CTA GCC ATT GAG
 Ala Val Leu Arg Trp Thr Arg Glu Lys Gln Pro Asn Tyr Gly Leu Ala Ile Glu

1122 1137 1152 1167
 GTG ACT CAC CTC CAT CAG ACT CGG ACC CAC CAG GGC CAG CAT GTC AGG ATT AGC
 Val Thr His Leu His Gln Thr Arg Thr His Gln Gly Gln His Val Arg Ile Ser

1182 1197 1212
 CGA TCG TTA CCT CAA GGG AGT GGG AAT TGG GCC CAG CTC CGG CCC CTC CTG GTC
 Arg Ser Leu Pro Gln Gly Ser Gly Asn Trp Ala Gln Leu Arg Pro Leu Leu Val

1227 1242 1257 1272
 ACC TTT GGC CAT GAT GGC CGG GGC CAT GCC TTG ACC CGA CGC CGG AGG GCC AAG
 Thr Phe Gly His Asp Gly Arg Gly His Ala Leu Thr Arg Arg Arg Arg Ala Lys

1287 1302 1317
 CGT AGC CCT AAG CAT CAC TCA CAG CGG GCC AGG AAG AAG AAT AAG AAC TGC CGC
 Arg Ser Pro Lys His His Ser Gln Arg Ala Arg Lys Lys Asn Lys Asn Cys Arg

1332(311) 1347 1362 1377
 CGC CAC TCG CTC TAT GTG GAC TTC AGC GAT GTG GGC TGG AAT GAC TGG ATT GTG
 Arg His Ser Leu Tyr Val Asp Phe Ser Asp Val Gly Trp Asn Asp Trp Ile Val

1392 1407 1422 1437
 GCC CCA CCA GGC TAC CAG GCC TTC TAC TGC CAT GGG GAC TGC CCC TTT CCA CTG
 Ala Pro Pro Gly Tyr Gln Ala Phe Tyr Cys His Gly Asp Cys Pro Phe Pro Leu

1452 1467 1482
 GCT GAC CAC CTC AAC TCA ACC AAC CAT GCC ATT GTG CAG ACC CTG GTC AAT TCT
 Ala Asp His Leu Asn Ser Thr Asn His Ala Ile Val Gln Thr Leu Val Asn Ser

1497 1512 1527 1542
 GTC AAT TCC AGT ATC CCC AAA GCC TGT TGT GTG CCC ACT GAA CTG AGT GCC ATC
 Val Asn Ser Ser Ile Pro Lys Ala Cys Cys Val Pro Thr Glu Leu Ser Ala Ile

Figure 2B

1557 1572 1587
 TCC ATG CTG TAC CTG GAT GAG TAT GAT AAG GTG GTA CTG AAA AAT TAT CAG GAG
 Ser MET Leu Tyr Leu Asp Glu Tyr Asp Lys Val Val Leu Lys Asn Tyr Gln Glu

1602 1617 (408) 1636 1646 1656
 ATG GTA GTA GAG GGA TGT GGG TGC CGC TGAGATCAGG CAGTCCTTGA GGATAGACAG
MET Val Val Glu Gly Cys Gly Cys Arg

1666 1676 1686 1696 1706 1716 1726
 ATATACACAC CACACACACA CACCACATAC ACCACACACA CACGTTCCCA TCCACTCACC CACACACTAC

1736 1746 1756 1766 1776 1786 1796
 ACAGACTGCT TCCTTATAGC TGGACTTTTA TTTAAAAAAA AAAAAAAAAA AATGGAAAAA ATCCCTAAAC

1806 1816 1826 1836 1846 1856 1866
 ATTCACCTTG ACCTTATTTA TGACTTTACG TGCAAATGTT TTGACCATAT TGATCATATA TTTTGACAAA

1876 1886 1896 1906 1916 1926 1936
 ATATATTTAT AACTACGTAT TAAAGAAAA AAATAAATG AGTCATTATT TTAAAAAAA AAAAAAACT

1946
 CTAGAGTCGA CGGAATTC

Figure 3

TGA ACA AGA GAG TGC TCA AGA AGC TGT CCA AGG ACG GCT CCA CAG AGG * Thr Arg Glu Cys Ser Arg Ser Cys Pro Arg Thr Ala Pro Gln Arg -41 -40 -35 -30	48
CAG GTG AGA GCA GTC ACG AGG AGG ACA CGG ATG GCG CAC GTG GCT GCG Gln Val Arg Ala Val Thr Arg Arg Thr Arg Met Ala His Val Ala Ala -25 -20 -15 -10	96
GGG TCG ACT TTA GCC AGG CGG AAA AGG AGC GCC GGG GCT GGC AGC CAC Gly Ser Thr Leu Ala Arg Arg Lys Arg Ser Ala Gly Ala Gly Ser His -5 1 5	144
TGT CAA AAG ACC TCC CTG CGG GTA AAC TTC GAG GAC ATC GGC TGG GAC Cys Gln Lys Thr Ser Leu Arg Val Asn Phe Glu Asp Ile Gly Trp Asp 10 15 20	192
AGC TGG ATC ATT GCA CCC AAG GAG TAT GAA GCC TAC GAG TGT AAG GGC Ser Trp Ile Ile Ala Pro Lys Glu Tyr Glu Ala Tyr Glu Cys Lys Gly 25 30 35	240
GGC TGC TTC TTC CCC TTG GCT GAC GAT GTG ACG CCG ACG AAA CAC GCT Gly Cys Phe Phe Pro Leu Ala Asp Asp Val Thr Pro Thr Lys His Ala 40 45 50 55	288
ATC GTG CAG ACC CTG GTG CAT CTC AAG TTC CCC ACA AAG GTG GGC AAG Ile Val Gln Thr Leu Val His Leu Lys Phe Pro Thr Lys Val Gly Lys 60 65 70	336
GCC TGC TGT GTG CCC ACC AAA CTG AGC CCC ATC TCC GTC CTC TAC AAG Ala Cys Cys Val Pro Thr Lys Leu Ser Pro Ile Ser Val Leu Tyr Lys 75 80 85	384
GAT GAC ATG GGG GTG CCC ACC CTC AAG TAC CAT TAC GAG GGC ATG AGC Asp Asp Met Gly Val Pro Thr Leu Lys Tyr His Tyr Glu Gly Met Ser 90 95 100	432
GTG GCA GAG TGT GGG TGC AGG TAGTATCTGC CTGCGGG Val Ala Glu Cys Gly Cys Arg 105 110	470

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